HATCHERY AND GENETIC MANAGEMENT PLAN (HGMP)

Hatchery Program:	Kalama River Tule Fall Chinook (Integrated)
Species or Hatchery Stock:	Fall Chinook (*Oncorhynchus tshawytscha*) Kalama River Stock
Agency/Operator:	Washington Department of Fish and Wildlife
Watershed and Region:	Kalama River/Lower Columbia
Date Submitted:	
Date Last Updated:	August 19, 2014

Executive Summary

The Washington Department of Fish and Wildlife is submitting a Hatchery and Genetic Management Plan (HGMP) for the Kalama River Tule Fall Chinook program to the National Marine Fisheries (NMFS) for consultation under Section 10(a)(1)(A) or 4(d) of the Endangered Species Act (ESA). NMFS will use the information in this HGMP to evaluate the hatchery impacts on salmon and steelhead listed under the ESA. The primary goal of an HGMP is to devise biologically-based hatchery management strategies that ensure the conservation and recovery of salmon and steelhead populations. This HGMP focuses on the implementation of hatchery reform actions adopted by the Washington Fish and Wildlife Commission Policy on Hatchery and Fishery Reform C-3619.

The purpose of the program is to produce Kalama River Tule Chinook for sustainable escapement to the watershed, while providing recreational fisheries. Program fish will be produced at the Kalama Falls Hatchery, located on the Kalama River (WRIA 27.0002), and Fallert Creek Hatchery, located on Hatchery/Fallert Creek (WRIA 27.0017, tributary to the Kalama River). The program will annually release 7,000,000 sub-yearlings to the Kalama River.

This fall Chinook HGMP is built around the principles and recommendations of the Hatchery Scientific Review Group (HSRG). These principles and recommendations represent the best science available for operating hatchery facilities consistent with the conservation of salmonid species. The program has been operated as a "integrated type" program, as defined by the HSRG, since 2010. An "integrated" program is one in which natural-origin individuals are used in the hatchery broodstocks. Integration is achieved by using up to 30% of the returning adult natural-origin fall Chinook (distinguished by an intact adipose fin) returning to the Kalama River at the Modrow Weir (RKm 4.8) and Fallert and Kalama Falls Hatchery traps (RKm 7.9 and 36.8, respectively) from August through mid-October. All fish released through this hatchery program have been 100% mass-marked (adipose fin-clipped) since 2005; return year 2010, of these, 250,000 sub-yearling fall Chinook are also released coded-wire tagged (CWT).

The Lower Columbia River Chinook are listed as "Threatened" under the ESA. The ESU includes the Kalama Tule Chinook Program.

Broodstock Collection:
The broodstock is derived from stock returning to the Kalama sub-basin. The proportion of natural-origin fish in the broodstock (pNOB) has averaged 10% over the last five years. The current egg-take goal is 7.7-million at Kalama Falls and Fallert Creek Hatcheries; up to 1,475 adult pairs may be collected. Surplus hatchery fish in excess of broodstock may be donated to food banks or used for nutrient enhancement.

Harvest:
Total annual harvest is dependent on management response to annual abundance in *Pacific Salmon Commission* (PSC - U.S./Canada), *Pacific Fishery Management Council* (PFMC - U.S. ocean), and *Columbia River Compact* forums. WDFW has also received authorization for tributary, Columbia River mainstem, and ocean fisheries; the combined harvest rates in the *Fisheries Management and Evaluation Plan* (FMEP), *Columbia River Fish Management Plan* (CRFMP), and ocean fisheries are reviewed annually in the North of Falcon process to ensure the harvest rates are consistent with recovery of the Lower Columbia River Tule Chinook population. The *U.S. v Oregon* Technical Advisory Committee (TAC) has prepared Biological Assessments (BAs) for combined fisheries based on relevant *U.S. v Oregon* management plans and agreements. The current BA concerns Columbia River treaty Indian and non-Indian fisheries, as described in the "2008–2017 *U.S. v Oregon* Management Agreement for upriver Chinook, sockeye, steelhead, coho, and white sturgeon" (2008–2017 MA).

Due to limitations that not all fish can be accounted for as being harvested or as back-to-rack counts, smolt-to-adult survival rates (SAR) are likely underestimated. Based on the average SAR of 0.265% for brood years 2000-2009, and a programmed release goal of 7,000,000 sub-yearlings, the estimated production goal would be 18,550 adults.

Monitoring and Evaluation:
Performance indicators for harvest will be accomplished by continuing mass-marking (adipose fin-clip); CWT recoveries help determine stray rate contributions on spawning grounds by watersheds close in proximity to this program's release vicinity.

In addition, temporary fish collection weirs were installed, and operated on the lower Grays, Coweeman, Kalama, Green, Washougal and Elochoman rivers. Operation of these weirs allow WDFW to control the number of hatchery fall Chinook reaching natural spawning locations, thereby benefiting natural production in these basins. Additionally, this project will fund spawning ground survey activities to monitor the effectiveness of these weirs and allow for the calculation of important hatchery performance metrics, such as pHOS. Deliverables include estimates of pHOS, and trapping efficiency, plus a draft Section 10 report for the weir on the Grays Elochoman and Coweeman Rivers.

Operation and Maintenance of Hatchery Facilities:
The Kalama Tule fall Chinook program uses two facilities. Kalama Falls Hatchery has water rights to divert water at a rate of 270 cfs from the Kalama River and 5 cfs from two non-fish bearing unnamed creeks. Fallert Creek Hatchery has water rights to divert water at a rate of 8.7 cfs from the Kalama River and 25 cfs from Fallert Creek. Kalama Falls Hatchery is a 100% barrier to fish passage, with a diversion dam forcing fish to enter a step and pool ladder leading to a concrete trapping area. Fallert Creek Hatchery prevents passage upstream into Fallert Creek; rebuild of a new intake system is expected to be completed by 2020. The return water systems operate under a National Pollutant Discharge Elimination System (NPDES) permit.

1 SECTION 1. GENERAL PROGRAM DESCRIPTION

1.1 Name of hatchery or program.

Kalama River Fall Chinook

1.2 Species and population (or stock) under propagation, and ESA status.

Kalama Fall Chinook (*Oncorhynchus tshawytscha*)

ESA Status: "Threatened" March 24, 1999 (64FR14308); reaffirmed on August 15, 2011 (76 FR 50448).

1.3 Responsible organization and individuals

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Other agencies, Tribes, co-operators, or organizations involved, including contractors, and extent of involvement in the program:

NOAA-National Marine Fisheries Service (NMFS) – Manager of Mitchell Act Funds

1.4 Funding source, staffing level, and annual hatchery program operational costs.

Funding Sources	Operation Information
Mitchell Act	Full time equivalent staff –7.0
	Annual operating cost (dollars) - $1,153,513

The above information for full-time equivalent staff and annual operating cost applies cumulatively to anadromous program facilities and cannot be broken out specifically by program.

1.5 Location(s) of hatchery and associated facilities.

Broodstock Source: Kalama River Fall Chinook

Table 1.5.1: Location of culturing phases, by facility.

Facility	Culturing Phase	Location
Modrow RB Weir	Broodstock collection	Kalama River (WRIA 27.0002) at RKm 4.8, at the Modrow Bridge on Modrow Road, Kalama WA
Kalama Falls Hatchery	Broodstock collection, Adult holding/spawning, Incubation, Rearing, Acclimation	Kalama River (WRIA 27.0002) at RM 36.8 (RKm 59.2); tributary to the Columbia River at RM 73.1 (RKm 117.6), Lower Columbia River, Washington.
Fallert Creek Hatchery	Broodstock collection, Adult holding/spawning,	Hatchery/Fallert Creek (WRIA 27.0017) at RKm 0.5; tributary to the Kalama River at RKm 7.9 (RM 4.9);

	Incubation, Rearing, Acclimation	Lower Columbia River, Washington.

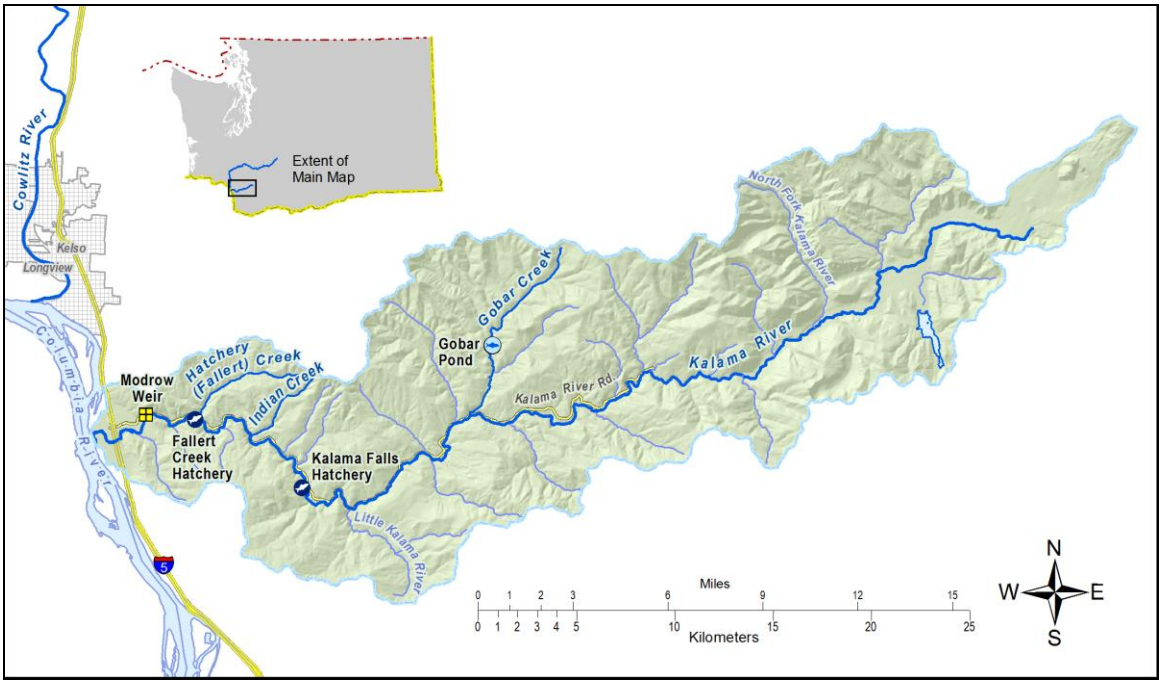

Figure 1.1: Map of Kalama Basin, including Modrow Weir, and Fallert Creek and Kalama Falls Hatchery. Source: WDFW GIS 2014.

1.6 Type of program.

Integrated Harvest

1.7 Purpose (Goal) of program.

Mitigation/Augmentation. The goal of this program is to provide escapement to the watershed and meet sport harvest goals under mark-selective fishery regulations (retention of adipose-clipped fish only), while minimizing impacts to natural-origin listed salmonids and steelhead. Also serves as mitigation for development (including hydro-power) and habitat degradation.

1.8 Justification for the program.

The program is funded through the Mitchell Act via NOAA-NMFS for the purpose of mitigation for lost fish production due to development within the Columbia River Basin.

WDFW protects listed fish and provides harvest opportunity on hatchery fish through the Lower Columbia River - *Fish Management and Evaluation Plan* (FMEP) (WDFW 2001). All mainstem and tributary fisheries are managed as mark-selective (no wild retention) fisheries to minimize the impact on listed wild fish.

To minimize impact on listed fish by the Kalama fall Chinook program and operations, the following risk aversions are included in this HGMP (**Table 1.8.1**).

Table 1.8.1: Summary of risk aversion measures for the Kalama Fall Chinook program.

Potential Hazard	HGMP Reference	Risk Aversion Measures
Water Withdrawal	4.1	Water rights are formalized through trust water right from the Department of Ecology. Monitoring and measurement of water usage is reported in monthly NPDES reports.
Intake Screening	4.1	*Kalama Falls Hatchery*. The new intake structure is

		compliant. *Fallert Creek Hatchery*. Intake screens at Fallert Creek are in compliance with state and federal guidelines (NMFS 1995, 1996), but do not meet the current *Anadromous Salmonid Passage Facility Design* criteria (NMFS 2011). A feasibility report for the river intake was funded in 2011 completed in 2012. WDFW is in the process of designing a new river intake system to meet NOAA-NMFS compliance (Mitchell Act Intake and Fish Passage Study Report 2003), and has included it in the 2013-2015 Capital Budget Request.
Effluent Discharge	4.1	These facilities operate under the "U*pland Fin-Fish Hatching and Rearing*" *National Pollution Discharge Elimination System* (NPDES) administered by the Washington Department of Ecology (DOE).
Broodstock Collection & Adult Passage	7.9	Adults are collected at Kalama Falls and Fallert Creek Hatcheries and the Modrow Trap (RKm 4.8). See HGMP section 7.2. All fish are mass-marked prior to release. Broodstock collection and sorting procedures can quickly identify listed non-target listed fish, and if encountered, released per protocol to minimize impact as determined by WDFW Region 5 staff.
Disease Transmission	7.9, 10.11	The *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006) and the *Fish Health Policy in the Columbia Basin* details hatchery practices and operations designed to stop the introduction and/or spread of any diseases within the Columbia Basin. Also, *Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries* (Fish Health Policy Chapter 5, IHOT 1995).
Competition & Predation	2.2.3, 10.11	Fish are released at a time, size and the system and life history stage to foster rapid migration to marine waters, and to allow juvenile listed fish to grow to a size that reduces potential for predation. Current risk aversions and future considerations are being reviewed and evaluated for further minimizing impacts to listed fish.

1.9 List of program "Performance Standards".

See HGMP section 1.10. Standards and indicators are referenced from Northwest Power Planning Council (NPPC) Artificial Production Review (APR) (NPPC 2001).

1.10 List of program "Performance Indicators", designated by "benefits" and "risks."

1.10.1 "Performance Indicators" addressing benefits.

Table 1.10.1: "Performance Indicators" addressing benefits.

Benefits		
Performance Standard	**Performance Indicator**	**Monitoring & Evaluation**
3.1.2 Program contributes to mitigation requirements. Program provides mitigation for lost fish production due to development within the Columbia River Basin.	Number of fish released by program returning, or caught, as applicable to given mitigation requirements.	Annually estimate survival and contribution for each brood year released. This program provides mitigation for lost fish

		production due to development within the Columbia River Basin and contributes to a meaningful harvest in sport and commercial fisheries.
3.1.3 Program addresses ESA responsibilities.	Program complies with Federal ESA-listed fish take authorizations for harvest and hatchery actions.	Hatchery program operation addresses ESA requirements through the development and review of this HGMP. HGMP updated and re-submitted to NOAA with significant changes or under permit agreement. Compliance with ESA is managed with sport fishery regulations that minimize impacts to ESA-listed fish and are monitored by WDFW law enforcement officers. The FMEP outlines anticipated encounter rates and expected mortality rates for these fisheries. Creel surveys are being implemented to verify. Natural populations are monitored annually to assess trends and compare with goals. HGMP updated and re-submitted to NOAA with significant changes or under permit agreement.
3.2.1 Fish produced for harvest are propagated and released in a manner enabling effective harvest, as described in all applicable fisheries management plans, while adequately minimizing by-catch of non-target species.	Annual number of fish produced by this program caught in all fisheries, including estimates of fish released.	Annually mass-mark hatchery releases to differentiate hatchery from natural-origin fish and record estimates of mark rate. The external mark enables mark-selective fisheries, which can reduce directed harvest mortality on natural-origin fish. Harvest is regulated to meet appropriate biological assessment criteria. Agencies monitor harvests to provide up-to-date information. Estimate survival and contribution to fisheries for each brood year released.
3.3.1. Artificial propagation program contributes to an increasing number of spawners returning to natural spawning areas.	Annual number of naturally-produced adults or redds on the spawning grounds or selected natural production index areas.	Annually monitor and report returns to the hatchery and spawning grounds.
3.3.2 Releases are sufficiently marked to allow statistically significant evaluation of program contribution to natural	Percentage of total hatchery releases are identifiable as hatchery-origin fish. Mass-mark (fin-clip, CWT, otolith-mark,	Annually monitor and report size, number, mass-mark quality (mark rate/tag rate) and date of all hatchery releases by mark

production, and to evaluate effects of the program on the local natural population.	other, etc., depending on species) production fish to identify them from naturally produced fish.	type. Annually sample returning fish for the mass-mark and CWT in fisheries and at the hatchery; monitor and report numbers of estimated hatchery (marked) and natural (unmarked) fish. Report CWT analysis to RMIS database.
3.4.1 Fish collected for broodstock are taken throughout the return or spawning period in proportions approximating the timing and age distribution of population from which broodstock is taken.	Temporal distribution of broodstock collection at point of collection.	Collect broodstock representatively and systematically throughout the return (August through October). Collect annual run timing, age and sex composition and spawning escapement timing data. Adhere to WDFW spawning guidelines (Seidel 1983; HSRG 2009).
3.5.5 Juveniles are released at fully-smolted stage to benefit juvenile to adult survival rates, and reduce the likelihood for residualism and negative ecological interactions with natural-origin fish.	Level of smoltification (size, appearance, behavior, etc.) at release compared to WDFW rearing and release guidelines. Release type (forced, volitional, or direct).	Monitor fish condition in the facilities throughout all rearing stages. Annually monitor and record size, number, and date of release.
3.6.1 The hatchery program uses standard scientific procedures to evaluate various aspects of artificial propagation.	Apply basic monitoring standards in the hatchery: food conversion rates, growth trajectories, mark/tag rate error, weight distribution (CV).	Collect annual run timing, age and sex composition data upon adult return. Annually record growth rates, mark rate and size at release and release dates. See also HGMP section 11 for program monitoring and evaluation.
3.8.3 Non-monetary societal benefits for which the program is designed are achieved.	Program is designed to help achieve the end goal of conserving and stabilizing natural salmon populations.	Long-term monitoring of system population will indicate success of program.

1.10.2 "Performance Indicators" addressing risks.

Table 1.10.2: **"Performance indicators" addressing risks.**

Risks		
Performance Standard	**Performance Indicator**	**Monitoring & Evaluation**
3.1.3 Program addresses ESA responsibilities	Program complies with Federal ESA-listed fish take authorizations for harvest and hatchery actions.	HGMP is updated to reflect any major changes in program and resubmitted to NOAA fisheries. Program risks have been addressed in this HGMP through best available science hatchery

		management actions. WDFW staff annually reviews Future Brood Document (FBD) for stock, size, number, date and location of releases from all production programs. Monitor and record juvenile hatchery fish size, number, date of release and mass-mark quality; monitor contribution of hatchery adult fish to fisheries and escapement.
3.2.1 Fish produced for harvest are produced and released in a manner enabling effective harvest, as described in all applicable fisheries management plans, while adequately minimizing by-catch of non-target species.	Number of marks released and estimated proportion of marks in out-migrant juveniles and returning adults on the spawning ground. Production fish are mass-marked (adipose fin-clip) to allow for their differentiation from naturally-produced fish	Monitor and record juvenile hatchery fish size, number, date of release and mass-mark (fin clips, tags, etc.) quality; monitor contribution of hatchery adult fish to fisheries and escapement. Harvest is regulated to meet appropriate biological assessment criteria. Agencies monitor harvests and hatchery escapements to provide up-to-date information.
3.2.2 Release groups are sufficiently marked in a manner consistent with information needs and protocols to enable determination of impacts to natural- and hatchery-origin fish in fisheries.	Percentage of total hatchery releases are identifiable as hatchery-origin fish. Mass-mark (adipose-fin clip, CWT, otolith-mark, etc., depending on species) produced fish to allow for their differentiation from naturally produced fish for selective fisheries.	Annually monitor and report size, number, date of release and mass-mark quality (adipose fin-clip rate) of all hatchery releases. Annually assess harvest of mass-marked hatchery fish based on CRC estimates and creel surveys.
3.3.2 Releases are sufficiently marked to allow statistically significant evaluation of program contribution to natural production and to evaluate effects of the program on the local natural population.	All hatchery production is identifiable in some manner (fin-marks, tags, otolith, etc.) consistent with information needs.	Annually monitor and record size, number, date of release and mass-mark quality (tag rate) of hatchery releases. Examine returning fish encountered for the mass-mark (CWT) at the hatchery and on spawning grounds. Annually record numbers of estimated hatchery (marked) and natural (unmarked).
3.4.1 Fish collected for broodstock are taken throughout the return or spawning period in proportions approximating the timing and age distribution of population from which broodstock is taken.	Temporal and age distribution of broodstock collected, compared to that of naturally-produced population at collection point.	Collect annual run timing, age and sex composition and return timing data.
3.4.3 Life history characteristics of the natural population do not change as a result of the hatchery	Life history characteristics are measured in adult and juvenile hatchery fish: return timing, age	Collect annual run timing, origin, and age and sex composition data.

program.	and sex composition, spawning timing, and size in returning hatchery adults; size, growth rates, and survival to release in juvenile production. Life history patterns of juvenile and adult NOR are stable.	Annually monitor and record juvenile hatchery fish size, growth rates, number released, mass-mark/tag data, survival-to-release rates, and date of release. Examine returning fish for the mass-mark (fin-clips, CWTs) at broodstock collection points and on the spawning grounds. Annually record and report numbers of estimated hatchery (marked) and natural (unmarked).
3.5.1 Patterns of genetic variation within and among natural populations do not change significantly as a result of artificial production.	Within and between populations, genetic structure is not affected by artificial production.	See HGMP section 11 for M&E information.
3.5.2 Collection of broodstock does not adversely impact the genetic diversity of the naturally-spawning population.	Total number of natural-origin spawners (if any) reaching the collection facility. Timing of collection compared to overall run timing.	All on-station hatchery releases are identifiable in some manner (fin-marks, tags, etc.). Collect annual run timing, origin, and age and sex composition data. CWT data reported to RMIS. Examine returning fish for the mass-mark (fin-clips, CWTs) at broodstock collection points and on the spawning grounds. Annually record and report numbers of estimated hatchery (marked) and natural (unmarked).
3.5.3 Hatchery-origin adults in natural production areas do not negatively affect the total natural spawning population.	The ratio of observed and/or estimated total numbers of artificially-produced fish on natural spawning grounds, to total number of naturally-produced fish (pHOS).	Examine returning fish for the mass-mark (fin-clips, CWTs) at weirs, broodstock collection points and on the spawning grounds.
3.5.4 Juveniles are released on-station, or after sufficient acclimation to maximize homing ability to intended return locations.	Location of release (on-station, acclimation pond, direct plant). Release type (forced, volitional or direct stream release). Proportion of adult returns to program's intended return location, compared to fisheries and artificial or natural production areas.	Annually record and report release information, including location, method and age class in hatchery data systems (WDFW Hatcheries Headquarters Database).
3.5.5 Juveniles are released at fully-smolted stage.	Level of smoltification at release. Release type (forced, volitional or direct).	Annually monitor and record size, number, date of release and release type.
3.7.1 Hatchery facilities are operated in compliance with all	Annual reports indicating levels of compliance with applicable	Pathologists from WDFW's Fish Health Section monitor program

applicable fish health guidelines and facility operation standards and protocols (IHOT, PNFHPC, *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State*).	standards and criteria. Periodic audits indicating level of compliance with applicable standards and criteria.	monthly. Exams performed at each life stage may include tests for virus, bacteria, parasites and/or pathological changes, as needed. See also Attachment 1 for pre-release Fish Health History. The program is operated consistent with the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006), *Fish Health Policy in the Columbia Basin*, and *Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries* (Fish Health Policy Chapter 5, IHOT 1995).
3.7.2 Effluent from hatchery facility will not detrimentally affect natural populations.	Discharge water quality compared to applicable water quality standards by NPDES permit. WDFW water right permit compliance.	Flow and discharge reported in monthly NPDES reports.
3.7.3 Water withdrawals and in-stream water diversion structures for artificial production facility operation will not prevent access to natural spawning areas, affect spawning behavior of natural populations, or impact juvenile rearing environment.	Water withdrawals compared to NMFS, USFWS and WDFW applicable passage and screening criteria for juveniles and adults.	Barrier and intake structure compliance assessed and needed fixes are prioritized.
3.7.4 Releases do not introduce pathogens not already existing in the local populations, and do not significantly increase the levels of existing pathogens. Follow the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, revised 2006).	Necropsies of fish to assess health, nutritional status, and culture conditions.	DFW Fish Health Section inspect adult broodstock yearly for pathogens and monitor juvenile fish on a monthly basis to assess health and detect potential disease problems. A fish health database will be maintained to identify trends in fish health and disease and implement fish health management plans based on findings.
	Release and/or transfer exams for pathogens and parasites.	Examine fish 1 to 6 weeks prior to transfer or release, in accordance with the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006).
	Inspection of adult broodstock for pathogens and parasites.	At spawning, lots of 60 adult broodstock are examined for pathogens.

	Inspection of off-station fish/eggs prior to transfer to hatchery for pathogens and parasites.	Controls of specific fish pathogens through eggs/fish movements are conducted in accordance to the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006).
3.7.6 Adult broodstock collection operation does not significantly alter spatial and temporal distribution of any naturally-produced population.	Spatial and temporal spawning distribution of natural populations above and below broodstock collection site is currently compared to historic distribution.	Trap is checked daily. Non-target listed fish, when encountered, are returned to the river.
3.7.7 Weir/trapping operations do not result in significant stress, injury or mortality in natural populations.	Mortality rates in trap. Pre-spawning mortality rates of captured fish in the hatchery and/or after release.	Traps checked daily. Annually record and report abundances and observations of natural-origin fish at hatchery facilities.
3.7.8 Predation by artificially produced fish on naturally – produced fish does not significantly reduce numbers of natural fish.	Hatchery juveniles are raised to smolt-size and released from the hatchery at a time that fosters rapid migration downstream.	Hatchery smolt release size and time are monitored to quantify/minimize predation effects on naturally-origin salmon and steelhead (Sharpe et al. 2008).
3.8.1 Cost of program operation does not exceed the net economic value of fisheries in dollars per fish for all fisheries targeting this population.	Total cost of operation.	Compare annual operational cost of program to calculated fishery contribution value (Wegge 2009).
3.8.2. Juvenile production costs are comparable to or less than other regional programs designed for similar objectives.	Total cost of program operation.	Annually monitor and report feed costs and fish health actions.

1.11 Expected size of program.

1.11.1 Proposed annual broodstock collection level (maximum number of adult fish).

Up to 1,445 adult pairs, not including jacks, are needed to achieve the established egg-take goal of 7.7-million (FBD 2014). This is based on an average fecundity of around 5,400 eggs/female, and a pre-spawning mortality of 5%.

1.11.2 Proposed annual fish release levels (maximum number) by life stage and location.

Table 1.11.2.1: Proposed annual fish release levels (maximum number) by life stage and location**.**

Age Class	Max. No.	Location	Major Watershed	Eco-Province
Sub-yearlings	7,000,000	Kalama River	Kalama Sub-Basin	Lower Columbia

Source: Future Brood Document 2014.

The sub-yearling release is split 50/50 between Kalama Falls and Fallert Creek Hatcheries.

1.12 Current program performance, including estimated smolt-to-adult survival rates, adult production levels, and escapement levels. Indicate the source of these data.

Table 1.12.1: Total combined Kalama Fall Chinook program releases and adult returns to the Kalama system, including Kalama and Fallert Creek Hatcheries and the Modrow Trap 2002-2013.

Year	Total Release	Hatchery/Trap Escapement*
2002	4,998,908	7,376
2003	4,927,266	7,121
2004	4,728,814	5,368
2005	5,206,992	4,918
2006	4,028,621	5,354
2007	4,518,777	5,217
2008	3,892,674	7,503
2009	5,526,656	9,224
2010	7,283,492	3,941
2011	5,927,622	12,888
2012	6,738,907	13,456
2013	6,679,346	16,118
Average	**5,371,506**	**8,207**

Source: WDFW Hatcheries Headquarters Database 2014.

*Includes fish passed upstream at Modrow that may have subsequently recruited to Kalama or Fallert after being passed upstream. Without unique marking it is currently not possible to track these passed fish individually to final disposition.

See **Table 3.3.1.1** for SAR calculation.

1.13 Date program started (years in operation), or is expected to start.

Kalama Falls Hatchery began operations in 1958. Fallert Creek Hatchery began operations in 1895.

1.14 Expected duration of program.

Program is on-going, with no plans for termination.

1.15 Watersheds targeted by program.

Kalama River (WRIA 27.0002/ Kalama Subbasin/ Lower Columbia Province.

1.16 Indicate alternative actions considered for attaining program goals, and reasons why those actions are not being proposed.

1.16.1 Brief Overview of Key Issues.

Fall Chinook in the Kalama River are collected at the Modrow weir (RM 2/RKm 4.8) and reared at Kalama Falls and Fallert Creek Hatcheries. This is an integrated program; fish are collected according to a run timing curve developed from previous years' data. Hatchery fall Chinook have been mass-marked since the 2005 brood and 2010 return year. Prior to this, the level natural-origin fish included in the broodstock was unknown. Smolts are released from their respective rearing facilities.

1.16.2 Potential Alternatives to the Current Program

Alternative 1: *Eliminate the program*: This action would reduce potential interaction with natural populations and eliminate potential impacts on other ESA-listed species. Currently this program supports ocean, mainstem and tributary fisheries, and is consistent with the mitigation requirements.

Alternative 2: Collect all adult broodstock at Fallert Creek Hatchery trap. WDFW currently collects broodstock for this program at the Modrow Trap and Kalama Falls Hatchery (KFH), according to a timing curve based on historical data. Collection of adults at the Fallert Creek trap is a possibility, but low flows, especially early in the season, could result in broodstock collection that is not integrated or representative.

Alternative 3: Modify release time or location, and/or reduce program size. The primary ecological risks include competition, predation, and disease transfer between hatchery and natural-origin fish. Of greatest concerned is competition between natural- and hatchery-origin fall Chinook. Data from other Chinook populations suggests that natural fall Chinook juvenile out-migration peaks in February or March, and continues through July. WDFW hatchery fall Chinook salmon are released in late-June, near the end of the natural-origin migration. Additionally, this program release goal of 7.0M sub-yearlings was determined by Mitchell Act production goals and the Pacific Salmon Treaty.

1.16.3 Potential Reforms and Investments

Reform/Investment 1: Address passage facilities at Kalama Falls. Fish passage at Kalama Falls is managed by the Kalama Falls fish barrier and fish ladder. This system is antiquated and needs to be modernized into a sorting, moving, and loading system that will use water in the conveyance of adult fish, and cause no harm to wild or hatchery fish. Currently, design work is being conducted to address these issues.

Reform/Investment 2: *Modernize Modrow Trap*. This trap facility has several issues related to unsafe handling of adult listed fish. In 2011, WDFW proposed a comprehensive re-design to the trap that will sort, and load fish with a water to water transfer method to cause no harm to hatchery or wild stocks. This redesign is set to take place in 2014. The picket spacing will also be modified to eliminate smaller fish from passing through.

Reform/Investment 3: *Modifiy intake screens at Fallert Creek to meet fish passage criteria.* The Kalama River water intake at Fallert Creek intake does not meet the current *Anadromous Salmonid Passage Facility Design* criteria (NMFS 2011). A feasibility report for the river intake was funded in 2011, and completed in 2012. WDFW is in the process of designing a new river intake system to meet NOAA-NMFS compliance (Mitchell Act Intake and Fish Passage Study Report 2003), and has included it in the 2013-2015 Capital Budget Request.

2 SECTION 2. PROGRAM EFFECTS ON NMFS ESA-LISTED SALMONID POPULATIONS. (USFWS ESA-Listed Salmonid Species and Non-Salmonid Species are addressed in Addendum A)

2.1 List all ESA permits or authorizations in hand for the hatchery program.

None currently. This HGMP is submitted to the NOAA Fisheries for ESA consultation and take prohibition exemption under ESA section 4(d) or 10.

2.2 Provide descriptions, status, and projected take actions and levels for NMFS ESA-listed natural populations in the target area.

2.2.1 Description of NMFS ESA-listed salmonid population(s) affected by the program.

- Identify the NMFS ESA-listed population(s) that will be directly affected by the program.

Lower Columbia River Chinook (*Oncorhynchus tshawytscha*). Listed as "threatened" on March 24, 1999 (64FR14308); threatened status reaffirmed on June 28, 2005 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

- Identify the NMFS ESA-listed population(s) that may be incidentally affected by the program.

Lower Columbia River coho (*Oncorhynchus kisutch*). Identified as a candidate species on June 25, 1995 (60FR38011). Listed as threatened on June 28, 2005 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

Lower Columbia River steelhead (*Oncorhynchus mykiss*). Listed as a threatened species on March 19, 1998 (63FR13347); threatened status reaffirmed on January 5, 2006 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

Columbia River chum salmon (*Oncorhynchus keta*). Listed as threatened on March 25, 1999 (64FR14507); threatened status reaffirmed on June 28, 2005 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

Kalama River eulachon (*Thaleichthys pacificus):* The Southern Distinct Population Segment (DPS) of Pacific eulachon was listed as *Threatened* under the ESA on May 17, 2010 (75 FR 13012).

2.2.2 Status of NMFS ESA-listed salmonid population(s) affected by the program.

- Describe the status of the listed natural population(s) relative to "critical" and "viable" population thresholds.

Lower Columbia River Chinook: In Washington, the LCR Chinook ESU includes all naturally spawned Chinook populations from the mouth of the Columbia to a transitional point between Washington and Oregon east of the Hood River and the White Salmon River, as well as fifteen artificial propagation programs. Excluded are Upper Columbia River bright hatchery stocks that spawn in the mainstem Columbia River below Bonneville Dam and in other tributaries upstream from the Sandy River to the Hood and White Salmon rivers (NMFS 2014 79FR20802).

Status: Today only two of 32 historical populations – the North Fork Lewis and Sandy late-fall populations – are considered viable. Most populations (26 out of 32) have a very low probability of persistence over the next 100 years, and some populations are extirpated, or nearly so. Five of the six strata fall significantly short of the Willamette-Lower Columbia Technical Recovery Team (WLC TRT) criteria for viability. One stratum – Cascade late fall – meets the WLC TRT criteria (Dornbush and Sihler 2013). Dam construction eliminated habitat for a number of populations leading to the extirpation of spring Chinook salmon populations in the Upper Cowlitz, Cispus, Tilton, North Fork Lewis , Big White Salmon rivers, and fall Chinook populations in the Upper Cowlitz and Big White Salmon rivers (SHIEER, NMFS 2004). Projects to allow access have been initiated in the Cowlitz and Lewis systems but these are not close to producing self-sustaining populations; Condit Dam on the Big White Salmon River was breached October 26, 2011. Based on the 2010 recovery plan analyses, all of the 14 Tule populations (**Table 2.2.2.1**) are considered very high risk except one that is considered at high risk. The modeling conducted in association with Tule harvest management suggests that three of the populations (Coweeman, Lewis and Washougal) are at a somewhat lower risk (LCFRB 2010).

Table 2.2.2.1: Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River Chinook populations.

		Baseline viability					Prod.	Abundance		
Population	Contribution	A&P	S	D	Net	Obj.	target	Historical	Baseline	Target
Coast Fall										
Grays/Chinook	Contributing[2]	VL	H	VL	VL[2]	M+	+500%	800	<50	1,000
Eloch/Skam [C]	Primary	VL	H	L	VL[2]	H	+150%	3,000	<50	1,500
Mill/Aber/Germ	Primary[1]	VL	H	L	VL[2]	H	+155%	2,500	50	900
Youngs Bay (OR)	Stabilizing	--[3]	--[3]	--[3]	L	L	--[3]	--[3]	--[3]	--[3]
Big Creek (OR) [C]	Contributing[1]	--[3]	--[3]	--[3]	VL	L	--[3]	--[3]	--[3]	--[3]
Clatskanie (OR)	Primary	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]
Scappoose (OR)	Primary[1]	--[3]	--[3]	--[3]	L	H	--[3]	--[3]	--[3]	--[3]
Cascade Fall										
Lower Cowlitz [C]	Contributing	VL	H	M	VL[2]	M+	+50%	24,000	500	3,000
Upper Cowlitz	Stabilizing	VL	VL	M	VL	VL	--	28,000	0	--
Toutle[C]	Primary[1]	VL	H	M	VL[2]	H+	+265%	11,000	<50	4,000
Coweeman [G]	Primary	VL	H	H	VL[2]	H+	+80%	3,500	100	900
Kalama	Contributing[2]	VL	H	M	VL[2]	M	+110%	2,700	<50	500
Lewis [G]	Primary	VL	H	H	VL[2]	H+	+280%	2,600	<50	1,500
Salmon	Stabilizing	VL	H	M	VL	VL	--	n/a	<50	--
Washougal	Primary	VL	H	M	VL[2]	H+	+190%	2,600	<50	1,200
Clackamas (OR) [C]	Contributing	--[3]	--[3]	--[3]	VL	M	--[3]	--[3]	--[3]	--[3]
Sandy (OR)	Contributing[1]	--[3]	--[3]	--[3]	VL	M	--[3]	--[3]	--[3]	--[3]
Cascade L Fall										
Lewis NF [C,G]	Primary	VH	H	H	VH[1]	VH	0%	23,000	7,300	7,300
Sandy (OR) [C,G]	Primary	--[3]	--[3]	--[3]	H	VH	--[3]	--[3]	--[3]	--[3]
Cascade Spring										
Upper Cowlitz [C,G]	Primary	VL	L	M	VL[2]	H+	>500%	22,000	300	1,800
Cispus [C,G]	Primary	VL	L	M	VL[2]	H+	>500%	7,800	150	1,800
Tilton	Stabilizing	VL	VL	VL	VL	VL	0%	5,400	<100	--
Toutle	Contributing	VL	H	L	VL	M	>500%	3,100	100	1,100
Kalama	Contributing[2]	VL	H	L	VL	L	>500%	4,900	100	300
Lewis NF [C]	Primary	VL	L	M	VL	H	>500%	15,700	300	1,500
Sandy (OR) [C,G]	Primary	--[3]	--[3]	--[3]	M	H	--[3]	--[3]	--[3]	--[3]
Gorge Fall										
L. Gorge (WA/OR)	Contributing	VL	M	L	VL[2]	M	>500%	n/a	<50	1,200
U. Gorge (WA/OR) [C]	Contributing[1]	VL	M	L	VL[2]	M	>500%	n/a	<50	1,200
White Salmon [C]	Contributing	VL	L	L	VL	M	>500%	n/a	<50	500
Hood (OR)	Primary[4]	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]
Gorge Spring										
White Salmon [C]	Contributing	VL	VL	VL	VL	L+	>500%	n/a	<50	500
Hood (OR)	Primary	--[3]	--[3]	--[3]	VL	VH	--[3]	--[3]	--[3]	--[3]

Source: LCFRB 2010.

L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.

[1] Increase relative to interim Plan.

[2] Reduction relative to interim Plan.

[3] Addressed in Oregon Management Unit plan.

[C] Designated as a historical core population by the TRT.

[G] Designated as a historical legacy population by the TRT.

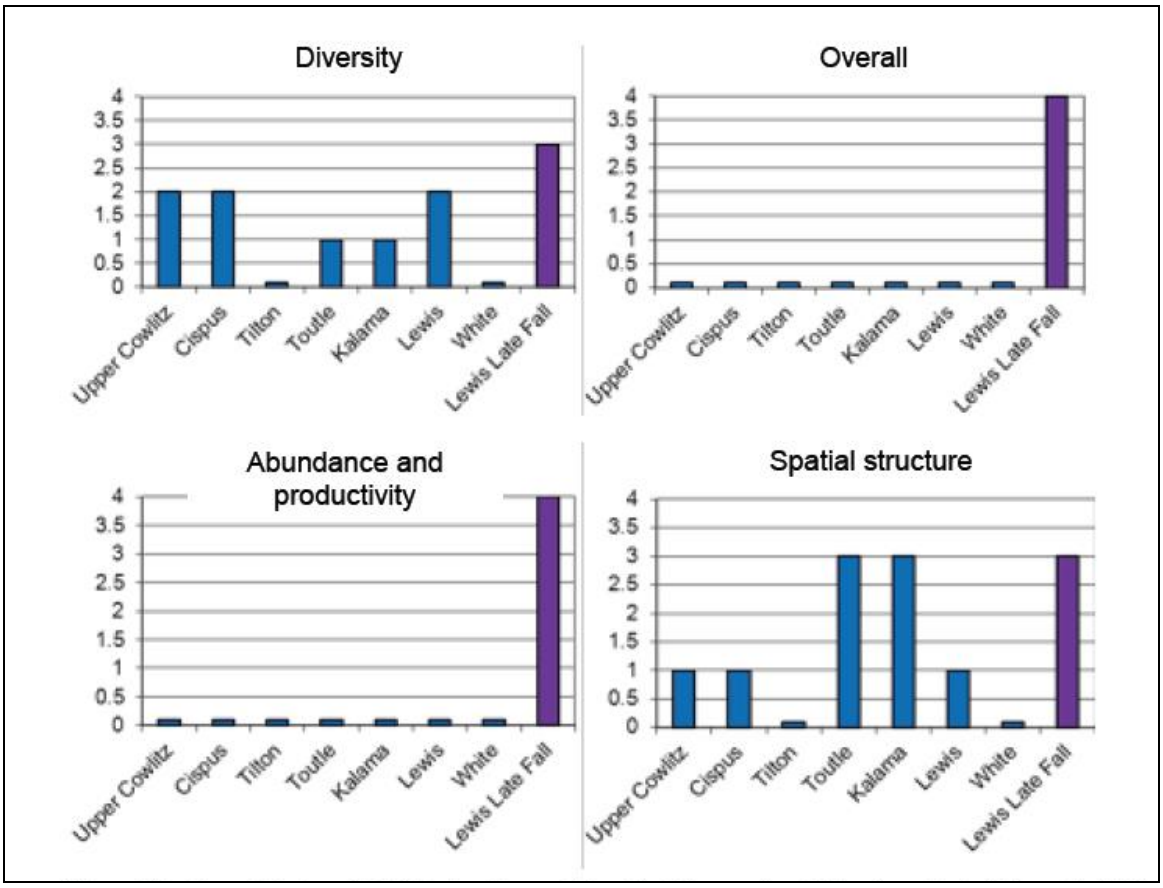

Figure 2.2.2.1: Current status of Washington lower Columbia River spring Chinook and late fall-run (bright) Chinook salmon populations for the VSP parameters and overall population risk. (LCFRB Recovery Plan 2010, chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

Lower Columbia River Steelhead *(Oncorhynchus mykiss)*: The DPS includes all naturally spawned anadromous *O. mykiss* (steelhead) populations below natural and manmade impassable barriers in streams and tributaries to the Columbia River between the Cowlitz and Wind Rivers, Washington (inclusive), and the Willamette and Hood Rivers, Oregon (inclusive), and excludes fish originating from the upper Willamette River Basin above Willamette Falls. The DPS includes seven artificial propagation programs, including the Cowlitz Trout Hatchery Winter-late (Lower Cowlitz), Kalama River Wild (winter- and summer-run) and Lewis River Wild Winter (NMFS 2014 79FR20802).

Status: Today, 16 of the 23 Lower Columbia River steelhead populations have a low or very low probability of persisting over the next 100 years, and six populations have a moderate probability of persistence. Only the summer-run Wind population is considered viable. All four strata in the DPS fall short of the WLC TRT criteria for viability (Dornbush and Sihler 2013). Populations in the upper Lewis and Cowlitz watersheds remain cut-off from access to essential spawning habitat by hydroelectric dams. Projects to allow access have been initiated in the Cowlitz and Lewis systems but these have not yet produced self-sustaining populations (Ford 2011). Condit Dam on the White Salmon River was breached October 26, 2011. WDFW is currently developing watershed-specific management plans in accordance with the SSMP. As part of this planning process, WDFW is proposing to complete a thorough review of current steelhead stock status using the most up to date estimates of adult abundance, juvenile production and genetic information.

Table 2.2.2.2: Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River steelhead populations.

		Baseline viability					Prod.	Abundance		
Population	Contribution	A&P	S	D	Net	Obj.	target	Historical	Baseline	Target
Coast Winter										
Grays/Chinook	Primary	VH	VH	M	M[1]	H	0%[4]	1,600	800	800
Eloch/Skam	Contributing	VH	VH	M	M[1]	M+	0%[4]	1,100	600	600
Mill/Ab/Germ	Primary	H	VH	M	M[1]	H	0%[4]	900	500	500
Youngs Bay (OR)	Primary	--[3]	--[3]	--[3]	VH	VH	--[3]	--[3]	--[3]	--[3]
Big Creek (OR)	Primary	--[3]	--[3]	--[3]	H	VH	--[3]	--[3]	--[3]	--[3]
Clatskanie (OR)	Primary	--[3]	--[3]	--[3]	VH	VH	--[3]	--[3]	--[3]	--[3]
Scappoose (OR)	Primary	--[3]	--[3]	--[3]	VH	VH	--[3]	--[3]	--[3]	--[3]
Cascade Winter										
Lower Cowlitz	Contributing	L	M	M	L	M	+5%	1,400	350	400
Upper Cowlitz [C,G]	Primary	VL	M	M	VL[2]	H[1]	>500%	1,400	<50	500
Cispus [C,G]	Primary	VL	M	M	VL[2]	H[1]	>500%	1,500	<50	500
Tilton	Contributing	VL	M	M	VL	L	>500%	1,700	<50	200
S.F. Toutle	Primary	M	VH	H	M	H+	+35%	3,600	350	600
N.F. Toutle [C]	Primary	VL	H	H	VL[2]	H	+125%		120	600
Coweeman	Primary	L	VH	VH	L[2]	H	+25%	900	350	500
Kalama	Primary	L	VH	H	L[2]	H+	+45%	800	300	600
N.F. Lewis [C]	Contributing	VL	M	M	VL[2]	M	>500%	8,300	150	400
E.F. Lewis	Primary	M	VH	M	M[1]	H	+25%	900	350	500
Salmon	Stabilizing	VL	H	M	VL[2]	VL	0%	na	<50	--
Washougal	Contributing	L	VH	M	L[2]	M	+15%	800	300	350
Clackamas (OR) [C]	Primary	--[3]	--[3]	--[3]	M	H	--[3]	--[3]	--[3]	--[3]
Sandy (OR) [C]	Primary	--[3]	--[3]	--[3]	L	VH	--[3]	--[3]	--[3]	--[3]
Cascade Summer										
Kalama [C]	Primary	H	VH	M	M[1]	H	0%[4]	1,000	500	500
N.F. Lewis	Stabilizing	VL	VL	VL	VL	VL	0%	na	150	--
E.F. Lewis [G]	Primary	VL	VH	M	VL[2]	H	>500%	600	<50	500
Washougal [C,G]	Primary	M	VH	M	M[1]	H	+40%	2,200	400	500
Gorge Winter										
L. Gorge (WA/OR)	Primary	L	VH	M	L[2]	H	+45%	na	200	300
U. Gorge (WA/OR)	Stabilizing	L	M	M	L[2]	L	0%	na	200	--
Hood (OR) [C,G]	Primary	--[3]	--[3]	--[3]	M	H	--[3]	--[3]	--[3]	--[3]
Gorge Summer										
Wind [C]	Primary	VH	VH	H	H[1]	VH	0%[4]	na	1,000	1,000
Hood (OR)	Primary	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]

Source: LCFRB 2010.

L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.

[1] Increase relative to interim Plan.

[2] Reduction relative to interim Plan.

[3] Addressed in Oregon Management Unit plan.

[4] Improvement increments are based on abundance and productivity; however, this population will require improvement in spatial structure or diversity to meet recovery objectives.

[C] Designated as a historical core population by the TRT.

[G] Designated as a historical legacy population by the TRT.

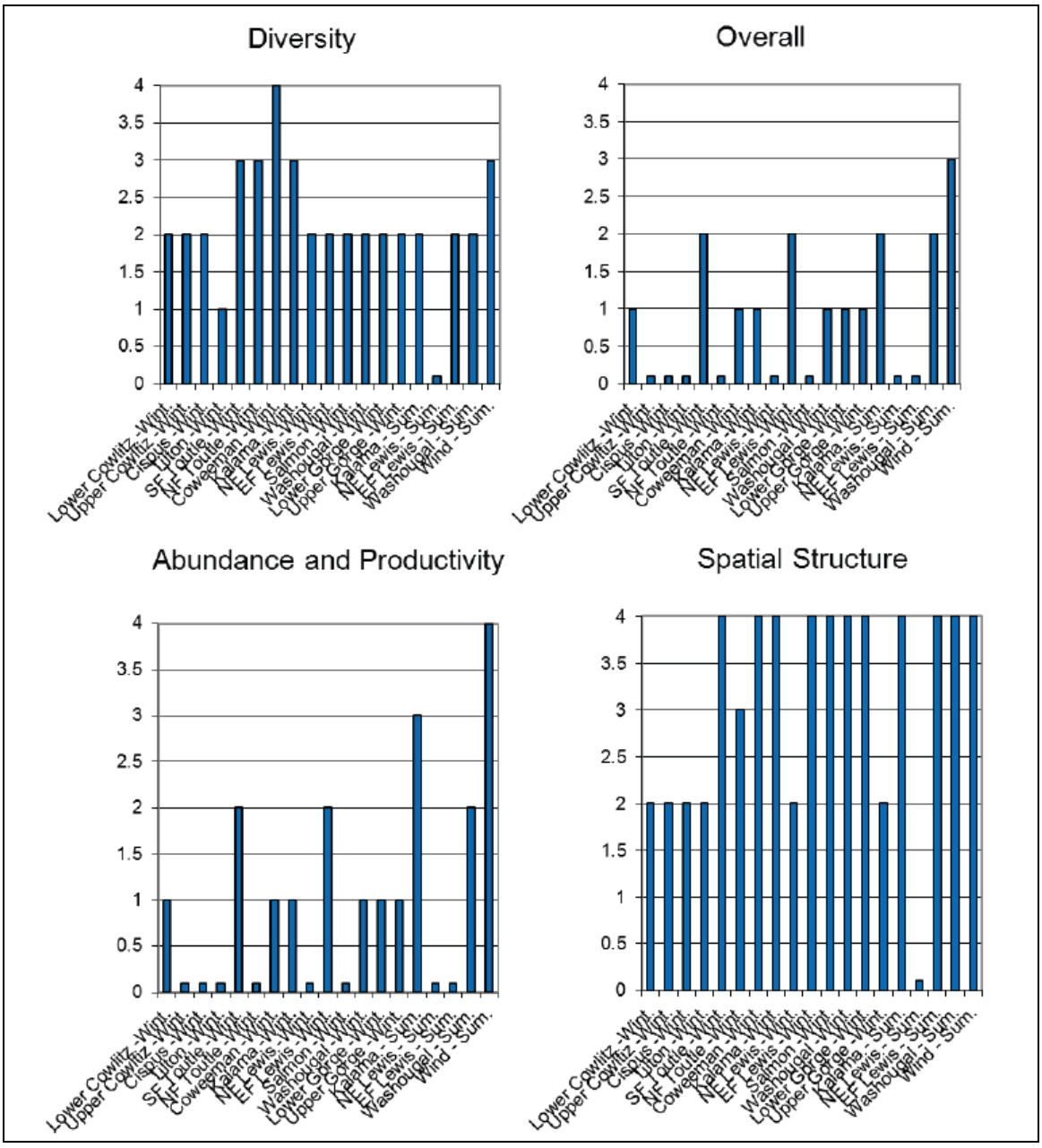

Figure 2.2.2.2: Current status of Washington LCR steelhead populations for the VSP parameters and overall population risk. (LCFRB 2010 Recovery Plan, chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

Lower Columbia River coho *(Oncorhynchus kisutch)*: Originally part of a larger Lower Columbia River/Southwest Washington ESU, Lower Columbia coho were identified as a separate ESU and listed as threatened on June 28, 2005. The ESU includes all naturally spawned populations of coho salmon in the Columbia River and its tributaries in Washington and Oregon, from the mouth of the Columbia up to and including the Big White Salmon and Hood Rivers. The twenty-one artificial propagation programs include: the Grays River, Peterson Coho Project, Cowlitz Type-N Coho Program in the Upper and Lower Cowlitz Rivers, Cowlitz Game and Anglers Coho Program, Friends of the Cowlitz Coho Program, North Fork Toutle River Hatchery, Kalama River Type-N and Type-S Coho Programs, Lewis River Type-N and Type-S

Coho programs, Fish First Wild Coho and Type-N Coho programs, Syverson Project Type-N Coho Program, and Washougal Hatchery Type-N Coho Program (NMFS 2014 79FR20802).

Status: Status evaluations of LCR coho status, all based on WLC-TRT criteria, have been conducted since the last BRT status update in 2005 (McElhany et al. 2007, Beamesderfer et al. 2010, LCFRB 2010, Dornbusch and Sihler 2013). All of these evaluations concluded that the ESU is currently at very high risk of extinction. All of the Washington side populations are considered at very high risk, although uncertainty is high because of a lack of adult spawner surveys. The 2005 BRT evaluation noted that smolt traps indicate some natural production in Washington populations, though given the high fraction of hatchery origin spawners suspected to occur in these populations it is not clear that any are self-sustaining (Ford 2011). Since this time WDFW has implemented an ESU wide monitoring program for LCR coho which began in 2010. Preliminary results indicate that natural origin population abundance may be higher than previously thought for certain populations (WDFW, unpublished). Results from the first 3 years of monitoring should be available in the near future. Currently, 21 of the 24 Lower Columbia River coho salmon populations are considered to have a very low probability of persisting over the next 100 years, and none is considered viable (Dornbusch and Sihler 2013). All three strata in the ESU fall significantly short of the WLC TRT criteria for viability.

Table 2.2.2.3: Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River coho populations.

		Baseline viability					Prod.	Abundance		
Population	Contribution	A&P	S	D	Net	Obj.	target	Historical	Baseline	Target
Coast										
Grays/Chinook [L]	Primary	VL	H	VL	VL[2]	H	+370%	3,800	<50	2,400
Eloch/Skam [L]	Primary	VL	H	VL	VL[2]	H	+170%	6,500	<50	2,400
Mill/Ab/Germ [L]	Contributing	VL	H	L	VL[2]	M	>500%	2,800	<50	1,800
Youngs (OR) [L]	Stabilizing	--[3]	--[3]	--[3]	VL	VL	--[3]	--[3]	--[3]	--[3]
Big Creek (OR) [L]	Stabilizing[2]	--[3]	--[3]	--[3]	VL	VL	--[3]	--[3]	--[3]	--[3]
Clatskanie (OR) [L]	Primary[1]	--[3]	--[3]	--[3]	L	VH	--[3]	--[3]	--[3]	--[3]
Scappoose (OR) [L]	Primary	--[3]	--[3]	--[3]	M	VH	--[3]	--[3]	--[3]	--[3]
Cascade										
Lower Cowlitz [L]	Primary	VL	M	M	VL[2]	H	+100%	18,000	500	3,700
Upper Cowlitz [E, L]	Primary[1]	VL	M	L	VL	H[1]	>500%	18,000	<50	2,000
Cispus [E, L]	Primary[1]	VL	M	L	VL	H[1]	>500%	8,000	<50	2,000
Tilton [E, L]	Stabilizing[2]	VL	M	L	VL	VL[2]	0%	5,600	<50	--
Toutle SF [E, L]	Primary	VL	H	M	VL[2]	H	+180%	27,000	<50	1,900
Toutle NF [E, L]	Primary	VL	M	L	VL[2]	H	+180%		<50	1,900
Coweeman [L]	Primary	VL	H	M	VL[2]	H	+170%	5,000	<50	1,200
Kalama [L]	Contributing	VL	H	L	VL[2]	L	>500%	800	<50	500
NF Lewis [E, L]	Contributing	VL	L	L	VL[2]	L	+50%	40,000	200	500
EF Lewis [E, L]	Primary	VL	H	M	VL[2]	H	>500%	3,000	<50	2,000
Salmon [L]	Stabilizing	VL	M	VL	VL	VL	0%	na	<50	--
Washougal [L]	Contributing	VL	H	L	VL[2]	M+	>500%	3,000	<50	1,500
Clackamas (OR) [E, L]	Primary	--[3]	--[3]	--[3]	M	VH	--[3]	--[3]	--[3]	--[3]
Sandy (OR) [E, L]	Primary	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]
Gorge										
L Gorge (WA/OR) [L]	Primary	VL	M	VL	VL[2]	H	+400%	na	<50	1,900
U Gorge (WA) [L]	Primary[1]	VL	M	VL	VL[2]	H	+400%	na	<50	1,900
U Gorge/Hood (OR)[E]	Contributing[4]	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]

Source: LCFRB 2010.
L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.
[1] Increase relative to interim Plan.
[2] Reduction relative to interim Plan.
[3] Addressed in Oregon Management Unit plan.
[4] Improvement increments are based on abundance and productivity; however, this population will require improvement in spatial structure or diversity to meet recovery objectives.
[E] Early run (Type S) coho stock.
[L] Late run (Type N) coho stock.
(Core and Legacy populations not designated by the TRT for coho).

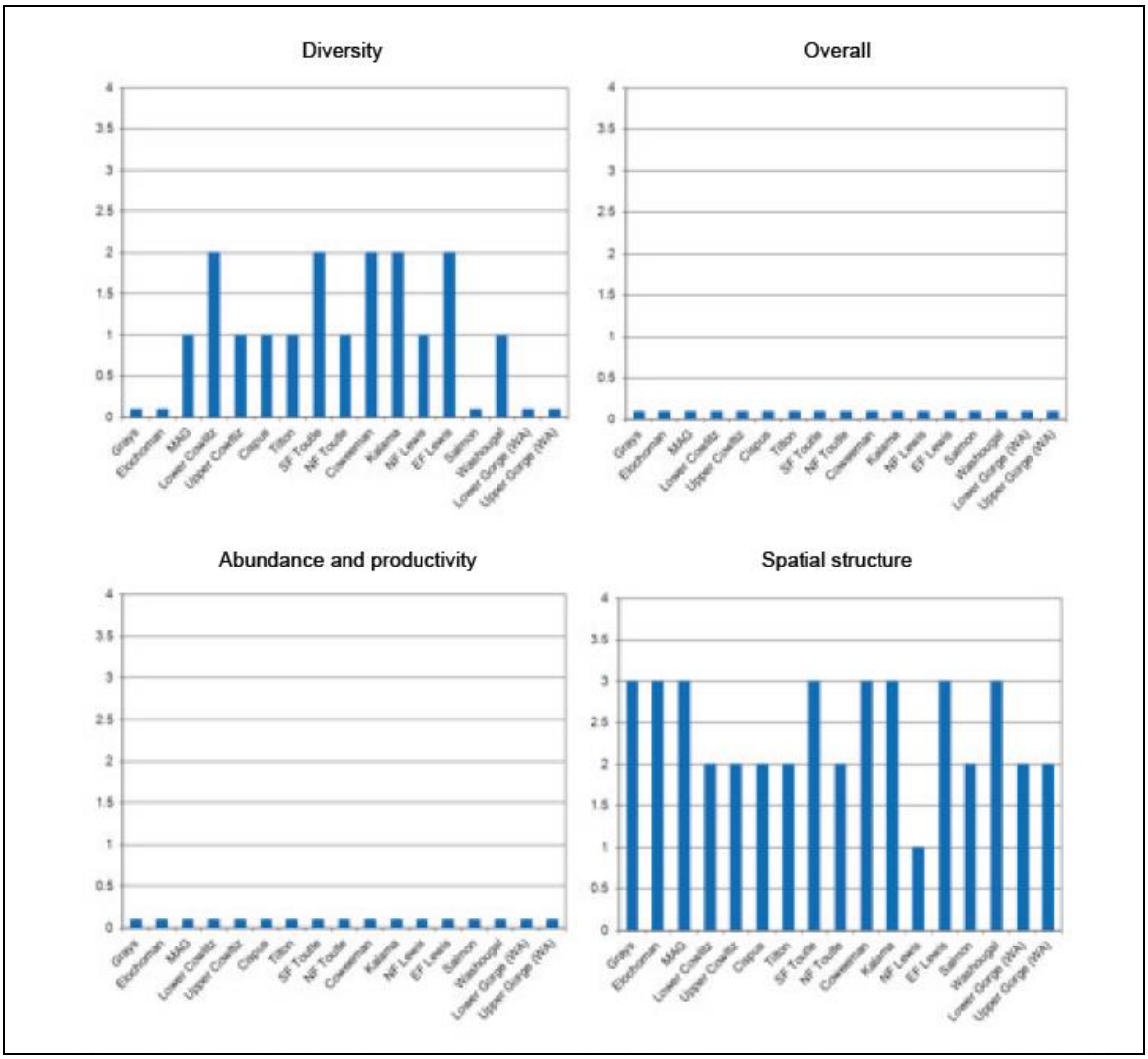

Figure 2.2.2.3: Current status of Washington LCR coho populations for the VSP parameters and overall population risk. (LCFRB 2010 recovery plan, chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

Columbia River chum salmon (*Oncorhynchus keta*). ESU includes all naturally spawned populations of chum salmon in the Columbia River and its tributaries in Washington and Oregon, as well as artificial propagation programs at Grays River and Washougal River/Duncan Creek chum hatchery programs (NMFS 2014 79FR20802).

Status: The LCFRB completed a revision recovery plan in 2010 that includes Washington populations of Columbia River chum salmon. This plan includes an assessment of the current status of Columbia River chum populations, which relied and built on the viability criteria developed by the WLC-TRT (McElhany et al. 2006) and an earlier evaluation of Oregon WLC populations (McElhany et al. 2007). This evaluation assessed the status of populations with regard to the VSP parameters of A/P, spatial structure, and diversity (McElhany et al. 2000). The result of this analysis is shown in **Figure 2.2.2.3**. The analysis indicates that all of the Washington populations with two exceptions are in the overall very high risk category (also described as extirpated or nearly so). The Grays River population was considered to be at moderate risk and the Lower Gorge population to be at low risk. The very high risk status assigned to the majority of Washington populations (and all the Oregon populations) reflects the very low abundance observed in these populations (e.g., <10 fish/year) (Ford 2011). Today, 15

of the 17 populations that historically made up this ESU are so depleted that either their baseline probability of persistence is very low or they are extirpated or nearly so; this is the case for all six of the Oregon populations. Currently almost all natural production occurs in just two populations: Grays/Chinook and the Lower Gorge. All three strata in the ESU fall significantly short of the WLC TRT criteria for viability (Dornbush and Sihler 2013).

Table 2.2.2.4: Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River chum populations.

		Baseline viability					Prod.	Abundance		
Population	Contribution	A&P	S	D	Net	Obj.	target	Historical	Baseline	Target
Coast										
Grays/Chinook [C,G]	Primary	VH	M	H	M[1]	VH	0%[4]	10,000	1,600	1,600
Eloch/Skam [C]	Primary	VL	H	L	VL[2]	H	>500%	16,000	<200	1,300
Mill/Ab/Germ	Primary	VL	H	L	VL	H	>500%	7,000	<100	1,300
Youngs (OR) [C]	Stabilizing[2]	--[3]	--[3]	--[3]	VL	VL	--[3]	--[3]	--[3]	--[3]
Big Creek (OR) [C]	Stabilizing[2]	--[3]	--[3]	--[3]	VL	VL	--[3]	--[3]	--[3]	--[3]
Clatskanie (OR)	Primary[1]	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]
Scappoose (OR)	Primary[1]	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]
Cascade										
Cowlitz (Fall) [C]	Contributing	VL	H	L	VL	M	>500%	195,000	<300	900
Cowlitz (Summer) [C]	Contributing	VL	L	L	VL	M	>500%	n/a	n/a	900
Kalama	Contributing	VL	H	L	VL	M	>500%	20,000	<100	900
Lewis [C]	Primary	VL	H	L	VL	H	>500%	125,000	<100	1,300
Salmon	Stabilizing	VL	L	L	VL	VL	0%	n/a	<100	--
Washougal	Primary	VL	H	L	VL[2]	H+	>500%	18,000	<100	1,300
Clackamas (OR) [C]	Contributing	--[3]	--[3]	--[3]	VL	M	--[3]	--[3]	--[3]	--[3]
Sandy (OR)	Primary	--[3]	--[3]	--[3]	VL	H	--[3]	--[3]	--[3]	--[3]
Gorge										
L. Gorge (WA/OR) [C,G]	Primary	VH	H	VH	H[1]	VH	0%[4]	6,000	2,000	2,000
U. Gorge (WA/OR)	Contributing	VL	L	L	VL	M	>500%	11,000	<50	900

Source: LCFRB 2010.
L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.
[5] Increase relative to interim Plan.
[6] Reduction relative to interim Plan.
[7] Addressed in Oregon Management Unit plan.
[8] Improvement increments are based on abundance and productivity; however, this population will require improvement in spatial structure or diversity to meet recovery objectives.
[C] Designated as a historical core population by the TRT.
[G] Designated as a historical legacy population by the TRT.

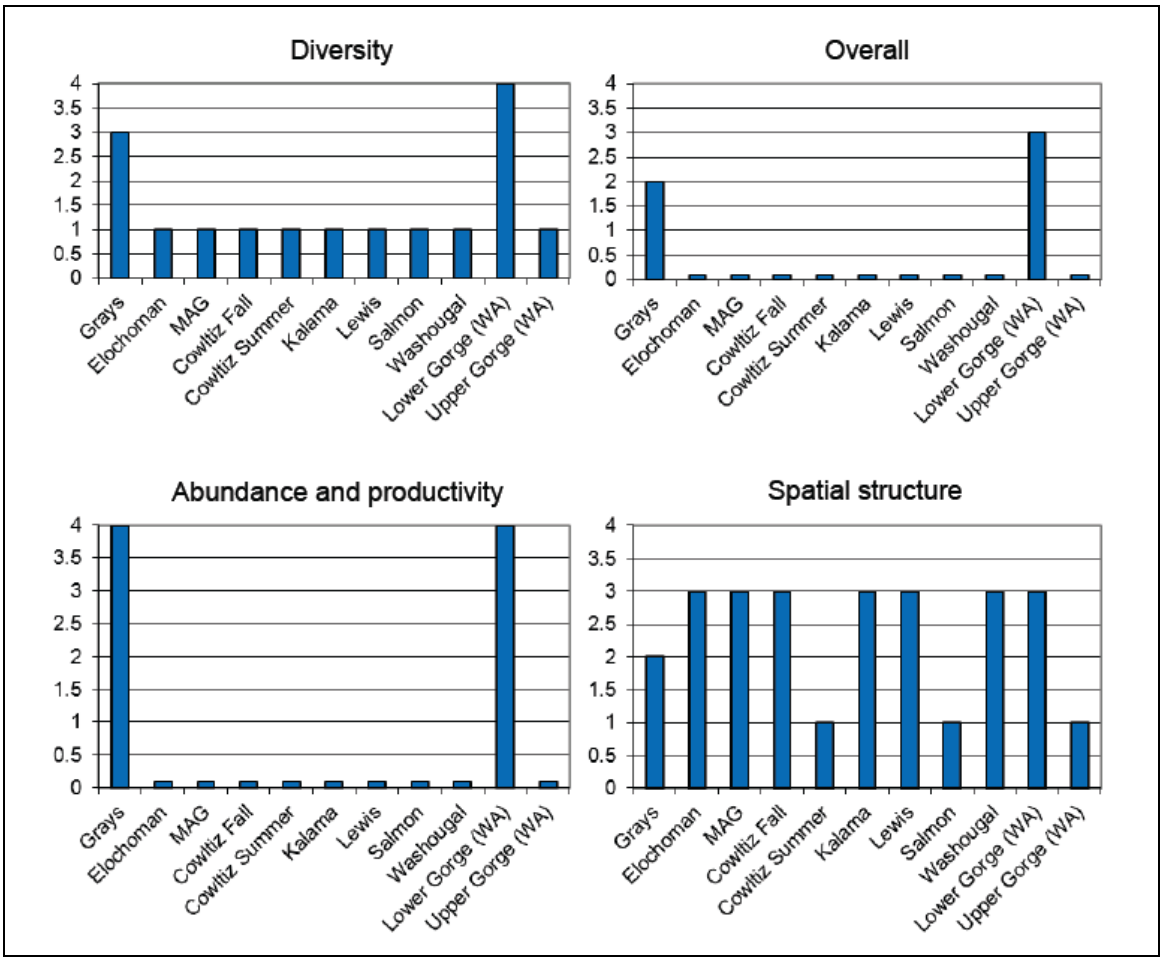

Figure 2.2.2.4: Current status of Washington CR chum populations for the VSP parameters and overall population risk. (LCFRB 2010 Recovery Plan, Chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

Kalama River eulachon (*Thaleichthys pacificus*): The Southern Distinct Population Segment (DPS) of Pacific eulachon was listed as *Threatened* under the ESA on May 17, 2010 (75 FR 13012).

Status: The lower Columbia River and its tributaries support the largest known spawning run of eulachon. The main stem of the lower Columbia River provides spawning and incubation sites, and major tributaries in Washington State that have supported runs in the past include the Grays, Elochoman, Cowlitz, Kalama and Lewis Rivers. Eulachon spawn in the Kalama River up to the confluence with Indian Creek and spawning has been confirmed as recently as 2011. The current abundance of eulachon is low and is declining in all surveyed populations throughout the DPS. The major threats and continued causes for declines in eulachon populations include climate change and its impacts on both ocean conditions and freshwater habitat, by-catch in commercial fisheries, dams and water diversions, degraded water quality, dredging and predation (NMFS 2011).

- Provide the most recent 12 year (e.g. 1988-present) progeny-to-parent ratios, survival data by life-stage, or other measures of productivity for the listed population.

Juvenile coho production estimates is the one measure of production in the Lower Columbia system. See HGMP section 11.1 for planned M&E.

Table 2.2.2.5: Lower Columbia River Washington tributary coho smolt production estimates, 1997-2009 (WDFW, Region 5).

Year	Cedar Creek	Mill Creek	Abernathy Creek	Germany Creek	Cowlitz Falls Dam	Mayfield Dam
1997	-----	-----	-----	-----	3,700	700
1998	38,400	-----	-----	-----	110,000	16,700
1999	28,000	-----	-----	-----	15,100	9,700
2000	20,300	-----	-----	-----	106,900	23,500
2001	24,200	6,300	6,500	8,200	334,700	82,200
2002	35,000	8,200	5,400	4,300	166,800	11,900
2003	36,700	10,500	9,600	6,200	403,600	38,900
2004	37,000	5,700	6,400	5,100	396,200	36,100
2005	58,300	11,400	9,000	4,900	766,100	40,900
2006	46,000	6,700	4,400	2,300	370,000	33,600
2007	29,300	7,000	3,300	2,300	277,400	34,200
2008	36,340	90,97	5,077	3,976	-----	38,917
2009	61,140	62,83	3,761	2,576	-----	29,718
2010	-----	-----	-----	-----	-----	49,171
2011	-----	-----	-----	-----	-----	43,831

Source: LCR FMEP Annual Report 2010 and WDFW Data 2012.

- Provide the most recent 12 year annual spawning abundance estimates, or any other abundance information. Indicate the source of these data.

Table 2.2.2.6: Spring Chinook salmon total spawner abundance estimates in LCR tributaries, 2000-2012.

Year	Cowlitz	Kalama	Lewis
2000	266	34	523
2001	347	578	754
2002	419	898	498
2003	1,953	790	745
2004	1,877	358	529
2005	405	380	122
2006	783	292	857
2007	74	2,150	264
2008	425	364	40
2009	763	34	80
2010	711	0	160
2011	1,359	26	120
2012	1,359	28	200

Source: Joe Hymer, WDFW Annual Database 2012.

Table 2.2.2.7: Fall Chinook salmon total spawner abundance estimates in LCR tributaries, 2000-2011[a].

Year	Elochoman River	Coweeman River[a]	Grays River	Skamokawa Creek	Cowlitz River	Green River (Toutle)	SF Toutle River	Kalama River	EF Lewis River	NF Lewis River	Washougal River
2000	884	424	80	482	2,100	1,580	204	3,877	391	6,504	2,757
2001	230	251	104	3	1,979	1,081	102	3,451	245	4,281	1,704
2002	332	566	390	7	3,038	5,654	216	10,560	441	5,518	2,728
2003	2,204	753	149	529	2,968	2,985	327	9,272	607	11,519	2,678
2004	4,796	1,590	745	2,109	4,621	4,188	618	6,680	918	13,987	10,597
2005	6,820	1,090	387	588	10,329	13,846	140	24,782	727	18,913	3,444
2006	7,581	900	82	372	14,427	7,477	450	18,952	1,375	17,106	6,050
2007	194	140	99	36	2,724	961	30	1,521	308	10,934	2,143
2008	782	95	311	253	1,334	824	45	2,617	236	4,268	3,182
2009	231	147	93	139	2,156	1,302	66	4,356	110	6,112	2,995
2010	1,883	1,330	12	268	2,762	605	NE	3,576	314	8,908	4,529
2011	508	2,148	353	41	1,616	668	NE	10,639	334	14,033	2,961

Source: Ron Roler, WDFW Natural Spawn Progress Reports 2012.
* Estimates of total adult and jack fall Chinook. May include fish put upstream of hatchery weirs.

Table 2.2.2.8: Wild winter steelhead escapement estimates for select SW Washington DPS populations, current WDFW escapement goals and LCSRP abundance targets.

Location	Grays River	Elochoman/ Skamokawa	Mill/Abernathy/ Germany
WDFW Escapement Goal	**1,486**	**853**	**508**
LCSRP Abundance Target	**800**	**600**	**500**
2000	1,064	650	380
2001	1,130	656	458
2002	724	370	354
2003	1,200	668	342
2004	1,132	768	446
2005	396	376	274
2006	718	632	398
2007	724	490	376
2008	764	666	528
2009	568	222	396
2010	422	534	398
2011	318	442	270
3-year average	**436**	**399**	**355**
5-year average	**559**	**471**	**394**
10-year average	**697**	**517**	**378**

Source: WDFW Data 2012

Table 2.2.2.9: Wild winter steelhead escapement estimates for select SW Washington DPS populations, current WDFW escapement goals and LCSRP abundance targets.

Location	Coweeman	SF Toutle	NF Toutle/ Green	Kalama	EF Lewis	Washougal
WDFW Escapement Goal	**1,064**	**1,058**	**NA**	**1,000**	**1,243**	**520**
LCSRP Abundance Target	**500**	**600**	**600**	**600**	**500**	**350**
2000	530	490	----	921	NA	NA
2001	384	348	----	1,042	377	216
2002	298	640	----	1,495	292	286
2003	460	1,510	----	1,815	532	764
2004	722	1,212	----	2,400	1,298	1,114
2005	370	520	388	1,856	246	320
2006	372	656	892	1,724	458	524
2007	384	548	565	1,050	448	632
2008	722	412	650	776	548	732
2009	602	498	699	1,044	688	418
2010	528	274	508	961	336	232
2011	408	210	416	622	308	204
3-year average	**513**	**327**	**541**	**876**	**444**	**285**
5-year average	**529**	**388**	**568**	**891**	**466**	**444**
10-year average	**487**	**648**	***588**	**1,374**	**515**	**523**

Source: WDFW Data 2012.
* 7-year average for NF Toutle/Green.

Table 2.2.2.10: Wild summer steelhead population estimates for LCR populations from 2001 to 2011, current WDFW escapement goals, and LCSRP abundance targets.

Location	Kalama	EF Lewis	Washougal	Wind
WDFW Escapement Goal	**1,000**	**NA**	**NA**	**1557**
LCSRP Abundance Target	**500**	**500**	**500**	**1,000**
2001	286	271	184	457
2002	454	440	404	680
2003	817	910	607	1096
2004	632	425	NA	861
2005	400	673	608	587
2006	387	560	636	632
2007	361	412	681	737
2008	237	365	755	614
2009	308	800	433	580
2010	370	602	787	788
2011	534	1,084*	956*	1,468
3-year average	**404**	**829**	**725**	**945**
5-year average	**362**	**653**	**722**	**837**
10-year average	**450**	**627**	**652**	**804**

Source: WDFW Data 2012.
* Preliminary estimates.

Table 2.2.2.11: Population estimates of chum salmon in the Columbia River.

Location	2002	2003	2004	2005	2006	2007	2008	2009	2010[a]	2011[a]
Crazy Johnson Creek	---	---	966	1,471	3,639	759	1,034	981	677	2,374
WF Grays River	---	---	9,015	1,324	1,232	1,909	800	994	1,967	7,002
Mainstem Grays River	---	---	4,872	1,400	1,244	1,164	886	750	3,467	1,848
I-205 area	3,468	2,844	2,102	1,009	862	544	626	1,132	2,105	4,947
Multnomah area	1,267	1,130	665	211	313	115	28	102	427	641
St Cloud area	---	137	104	92	173	9	1	14	99	509
Horsetail area	---	---	106	40	63	17	33	6	45	183
Ives area[b]	4,466	1,942	363	263	387	145	168	141	214	162
Duncan Creek[c]	13	16	2	7	42	9	2	26	48	85
Hardy Creek	343	392	49	73	104	14	3	39	137	173
Hamilton Creek	1,000	500	222	174	246	79	114	115	247	517
Hamilton Spring Channel	794	363	346	84	236	44	109	91	187	324
Grays return[d]	12,041	16,974	15,157	4,327	6,232	3,966	2,807	2,833	6,399	11,518
I-205 to Bonneville return	11,351	7,324	3,959	1,953	2,426	976	1,084	1,666	3,509	7,541
Lower Columbia River Total	23,392	24,298	19,116	6,280	8,658	4,942	3,891	4,499	9,908	19,059

Source: Todd Hillson - WDFW Chum Program 2012

[a] Data for 2010 and 2011 is preliminary.

[b] Ives area counts are the carcass tagging estimate plus fish removed for broodstock, except for 2007 and 2008, which is area under the curve.

[c] Totals for Duncan Creek do not include broodstock brought in from mainstem spawning areas, adult trap catch or surveys below monitoring weirs only..

[d] Grays return totals include natural spawners and removed for broodstock.

- Provide the most recent 12 year (e.g. 1988-1999) estimates of annual proportions of direct hatchery-origin and listed natural-origin fish on natural spawning grounds, if known.

The proportion of hatchery-origin spawners (pHOS) should be less than 30% of the naturally spawning population for this integrated program per HSRG guidelines (2009), as it is associated with a Contributing natural population. The estimated average pHOS based on CWT expansion data from 2010-2013 is 91%. See **Table 6.2.3.1** for the annually reported values.

2.2.3 Describe hatchery activities, including associated monitoring and evaluation and research programs, that may lead to the take of NMFS listed fish in the target area, and provide estimated annual levels of take.

- Describe hatchery activities that may lead to the take of listed salmonid populations in the target area, including how, where, and when the takes may occur, the risk potential for their occurrence, and the likely effects of the take.

Broodstock Program:

Broodstock Collection: Broodstock is collected at the Modrow Trap, located at approximately RKm 4.8, and the Kalama Falls and Fallert Creek Hatcheries. The Modrow Trap consists of a temporary rack-picket structure with a "V"-trap for capturing/holding adults. The trap structure is operated in the river during the August October period. Adults are sorted, and selected fish are transferred to a tanker truck via a brail-hoist system and transported to the Kalama Falls Hatchery. See also "take" tables at the end of this HGMP.

Genetic introgression: All fall Chinook adults recruited for broodstock have been from Kalama River returns. WDFW shifted to an integrated program, incorporating natural fish into the broodstock, in 2010.

No genetic analysis has been done on naturally-spawning Kalama fall Chinook. Allozyme analysis of Kalama hatchery fall chinook sampled in 1988 and 1989 showed that they were genetically

distinct from most other lower Columbia Tule fall Chinook but not significantly different from Abernathy Creek fall Chinook (Myers et al. 2006). Indirect take from genetic introgression is unknown.

Rearing Program:

Operation of Hatchery Facilities: Facility operation impacts include water withdrawal, effluent, and intake compliance. Effluent at outfall areas is rapidly diluted with mainstem flows and operation is within permitted NPDES guidelines (see HGMP sections 4.1 and 4.2). Indirect take from this operation is unknown.

Disease: Over the years, rearing densities, disease prevention and fish health monitoring have greatly improved the health of the hatchery programs. *Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries*-Chapter 5 (IHOT 1995) have been instrumental in reducing disease outbreaks. Although pathogens occur in the wild and fish might be affected, they are believed to go undetected with predation quickly removing those fish.

In addition, although pathogens may cause post release mortality in fish from hatcheries, there is little evidence that hatchery origin fish routinely infect natural populations of salmon and steelhead in the Pacific Northwest (Enhancement Planning Team 1986 and Steward and Bjornn 1990). Prior to release, the hatchery population health and condition is established by the Area Fish Health Specialist. This is commonly done one to three weeks pre-release, and up to six weeks on systems with pathogen-free water and little or no history of disease. Indirect take from disease is unknown.

Release:

Hatchery Production/Density-Dependent Effects: Hatcheries can release numbers of fish that can exceed the density of the natural productivity in a limited area for a short period of time and can compete with listed fish. Hatchery fish are released as active smolts that will emigrate quickly from the system. In addition, fish are released from both hatchery sites over a period of ten days to two weeks in order to minimize density effects. This strategy allows groups to emigrate and move from the area daily. Because Fallert Creek Hatchery is only 6.0 RKm upstream from the confluence with the Columbia, many of these hatchery Chinook smolts can vacate the Kalama system within the day that they are released. Indirect take from density dependent effects is unknown.

Potential Kalama Falls Hatchery fall Chinook predation and competition effects on listed salmonids and eulachon: The proposed annual production goal for this program is 7-million sub-yearlings. Chinook are released at 80 fpp (88 mm fl) in June (see HGMP section 10.3). Due to size differences between listed yearling and sub-yearling smolts (**Table 2.2.3.1**), competition is unlikely, with different prey items and habitat preferences. The impact of Fallert Creek releases on natural-origin salmon and steelhead would be minimized because the release location is below the majority of all known spawning sites for these populations in the Kalama River.

Table 2.2.3.1: Peak migration timing and average fork length (mm) of out-migrant juvenile Chinook, coho and steelhead captured in rotary screw traps on Mill, Germany and Abernathy creek, Lower Columbia River, 2008.

Stream	Chinook		Coho		Steelhead	
	Avg Size (mm)	Peak Migration	Avg Size (mm)	Peak Migration	Avg Size (mm)	Peak Migration
Mill Cr	37.0	Mar 10-Apr 13	104.2	Mar 17-23	154.5	Apr 28-May 4
Germany Cr	39.8	Mar 17-23	115.3	May 19-25	177.8	May 12-18
Abernathy Cr	37.9	Mar 31 – Apr 6	112.1	May 19-25	163.8	May 12-18

Source: Kinsel et al 2009.

Both juvenile and adult salmonids have been documented to feed on eulachon (Gustafson et al. 2010). Predation of eulachon by Chinook reared in this program may occur, however it is unknown to what degree such predation may occur.

Residualism: To maximize smolting characteristics and minimize residualism, WDFW adheres to a combination of acclimation, volitional release strategies, size, and time guidelines.

- Condition factors, standard deviation and co-efficient of variation (CV) are measured throughout the rearing cycle and at release.
- Feeding rates and regimes throughout the rearing cycle are programmed to satiation feeding to minimize out-of-size fish and programmed to produce smolt size fish at date of release.
- Based on past history, fish have reached a size and condition that indicates a smolted condition at release.
- Releases occur within known time periods of species emigration from acclimated ponds.
- Releases from these ponds are volitional with large proportions of the populations moving out initially with the remainder of the population vacating within days or a few weeks.
- Minimal residualism from WDFW Chinook programs following these guidelines has been indicated from snorkeling studies on the Elochoman River (Fuss et al. 2000) and on Nemah and Forks Creek (Riley et al. 2004). In extensive surveys conducted on the Lewis River, Hawkins and Tipping (1999) found no residualized hatchery Chinook. Indirect take from residualism is unknown.

Monitoring:

Associated monitoring Activities:

WDFW has implemented an expanded monitoring program for Chinook, coho, chum and steelhead populations in the Lower Columbia River (LCR) region of Southwest Washington (WDFW's Region 5) and fishery monitoring in the lower mainstem of the Columbia River. The focus of this expanded monitoring is to 1) gather data on Viable Salmonid Population (VSP) parameters – spawner abundance, including proportion of hatchery-origin spawners (pHOS), spatial distribution, diversity, and productivity, 2) to increase the coded wire tag (CWT) recovery rate from spawning grounds to meet regional standards, and 3) to evaluate the use of PIT tags to develop harvest rates for salmon and steelhead populations. Additionally, key watersheds are monitored for juvenile salmonid out-migrant abundance. Coupled with adult abundance information, these data sets allow for evaluation of freshwater productivity and development of biological reference points, such as seeding capacity. Monitoring protocols and analysis methods utilized are intended to produce unbiased estimates with measurements of precision in an effort to meet NOAA monitoring guidelines (Crawford and Rumsey 2009).

- Provide information regarding past takes associated with the hatchery program, (if known) including numbers taken, and observed injury or mortality levels for listed fish.

Table 2.2.3.2: Disposition of unmarked (no adipose fin-clip) fall Chinook returning to Kalama River facilities.

Brood Year	Mortality	Spawned
2010	24	104
2011	93	385
2012	31	192
2013	91	464

Source: WDFW Hatcheries Headquarters Database 2014.

- Provide projected annual take levels for listed fish by life stage (juvenile and adult) quantified (to the extent feasible) by the type of take resulting from the hatchery program (e.g. capture, handling, tagging, injury, or lethal take).

See "take" tables at the end of this HGMP.

- Indicate contingency plans for addressing situations where take levels within a given year have exceeded, or are projected to exceed, take levels described in this plan for the program.

No situations are expected to occur where take would exceed ESA limits. If significant numbers of wild salmonids are observed impacted by this operation, then staff would inform the WDFW District Biologist, Fish Health Specialist or Area Habitat Biologist who, along with the Hatchery Complex Manager, would determine an appropriate plan and consult with NOAA-NMFS for adaptive management review and protocols.

Handling and release of wild fall Chinook above broodstock needs is monitored. Any additional mortality from this operation on a yearly basis would be communicated to Fish Program staff for additional guidance.

3 SECTION 3. RELATIONSHIP OF PROGRAM TO OTHER MANAGEMENT OBJECTIVES

3.1 Describe alignment of the hatchery program with any ESU-wide hatchery plan (e.g. Hood Canal Summer Chum Conservation Initiative) or other regionally accepted policies (e.g. the NPPC Annual Production Review Report and Recommendations - NPPC document 99-15). Explain any proposed deviations from the plan or policies.

WDFW has several policies/plans that help inform management decisions regarding the HGMPs currently under review. These policies include:

1. Hatchery and Fishery Reform Policy (Commission Policy C3619)
2. The Conservation and Sustainable Fisheries Plan (draft)
3. The Hatchery Action Implementation Plans (HAIP)
4. Lower Columbia Salmon Recovery Plan (LCSRP)

Descriptions of these policies and excerpts are shown below:

Policies/Plans – Key Excerpts

Hatchery and Fishery Reform Policy: Washington Department of Fish and Wildlife Commission Policy C-3619. WDFW adopted the Hatchery and Fishery Reform Policy C-3619 in 2009. Its purpose is to advance the conservation and recovery of wild salmon and steelhead by promoting and guiding the implementation of hatchery reform. The intent of hatchery reform is to improve hatchery effectiveness, ensure compatibility between hatchery production and salmon recovery plans and rebuilding programs, and support sustainable fisheries. WDFW Policy C-3619 works to promote the conservation and recovery of wild salmon and steelhead and provide fishery-related benefits by establishing clear goals for each state hatchery, conducting scientifically defensible-operations, and using informed decision making to improve management. It is recognized that many state operated hatcheries are subject to provisions under *U.S. v Washington* (1974) and *U.S. v Oregon* and that hatchery reform actions must be done in close coordination with tribal co-managers. Washington Fish and Wildlife Commission Policy: POL-C3619.

Guidelines from the policy include:

1. Use the principles, standards, and recommendations of the Hatchery Scientific Review Group (HSRG) to guide the management of hatcheries operated by the Department.

2. Develop watershed-specific action plans that systematically implement hatchery reform as part of a comprehensive, integrated (All-H) strategy for meeting conservation and harvest goals at the watershed and Evolutionarily Significant Unit (ESU)/Distinct Population Segment (DPS) levels. Action Plans will include development of stock (watershed) specific population designations and application of HSRG broodstock management standards.

Conservation and Sustainable Fisheries Plan (*CSFP*)*:* The CSFP is a draft plan that has been developed to meet WDFW's responsibilities outlined in the Lower Columbia Salmon Recovery Plan (LCSRP) and address the HSRG suggested solutions and achieve HRSG standards for primary, contributing and stabilizing populations. The plan describes the implementation of changes to hatchery and harvest programs and how they assist in recovery and achieve HSRG guidelines. The draft plan also identifies Viable Salmonid Population (VSP) parameters that will be addressed.

Hatchery Action Implementation Plans (*HAIP*)*:* The HAIPs illustrate how WDFW is implementing hatchery programs to incorporate the HSRG guidelines. The plans provide the current programs and explain the future goals.

Lower Columbia Salmon Recovery Plan (*LCSRP*)*:* Some sub-basins will be free of hatchery influence and hatchery programs. In other sub-basins, hatchery programs will serve specific conservation and harvest purposes consistent with goals for naturally-spawning populations. The mosaic of programs is designed to ensure that overall each DPS will be naturally self-sustaining.

Strategies:

1. Reconfigure production-based hatchery programs to minimize impacts on natural populations and complement recovery objectives.
2. Adaptively manage hatcheries to respond to future knowledge, enhance natural production, and improve operational efficiencies.

Mitchell Act: This program receives Mitchell Act Funding. Initially passed in 1938, the Mitchell Act is intended to help rebuild and conserve the fish runs, and mitigate the impacts to fish from water diversions, dams on the mainstem of the Columbia River, pollution and logging. The Mitchell Act specifically directs establishment of salmon hatcheries, conduct of engineering and biological surveys and experiments, and installing fish protective devices. It also authorizes agreements with State fishery agencies and construction of facilities on State-owned lands. NMFS has administered the program as of 1970. There are 15 Mitchell Act hatcheries in Washington State; the majority of which are below Bonneville Dam.

The Mitchell Act programs are intended to support Northwest fishing economies – particularly coastal and Native American -- that have relied on Columbia River production both before and after dam construction. Catches of hatchery fish sustain the economies of local communities while keeping incidental mortalities of ESA-Listed fish at approved levels. Value of hatchery production and benefit to local economies will be further increased by implementing fisheries that increase harvest of hatchery produced fish, as expected through implementation of the LCSRP.

3.2 List all existing cooperative agreements, memoranda of understanding, memoranda of agreement, or other management plans or court orders under which program operates.

Future Brood Document. Hatchery salmon and steelhead production levels are detailed in the annual Future Brood Document, a pre-season planning document for fish hatchery production in Washington State for the upcoming brood stock collection and fish rearing season (July 1 – June 30).

See also HGMP section 3.1.

3.3 Relationship to harvest objectives.

Total annual harvest is dependent on management response to annual abundance in Pacific Salmon Commission (PSC - U.S./Canada), Pacific Fishery Management Council (PFMC - U.S. ocean), and Columbia River Compact forums. WDFW also has received authorization for tributary, Columbia River mainstem, and ocean fisheries; the combined harvest rates in the *Fisheries Management and Evaluation Plan* (FMEP), *Columbia River Fish Management Plan* (CRFMP), and ocean fisheries are reviewed annually in the North of Falcon process.

LCFRB Recovery Plan 2010. This near-term strategy involves limiting fishery impacts on natural populations to ameliorate extinction risks until a combination of measures can restore fishable natural populations. There is no directed Columbia River or tributary harvest of ESA-listed Kalama River salmon and steelhead. This practice will continue until the populations are sufficiently recovered to withstand such pressure and remain self-sustaining.

3.3.1 Describe fisheries benefitting from the program, and indicate harvest levels and rates for program-origin fish for the last twelve years (1988-99), if available.

Lower Columbia Chinook ESU consists of spring, fall Tule, and fall bright fish runs. These runs are impacted differently by fisheries outside the LCMA and outside WDFW management. The freshwater sport fishery occurs from August through October in the ten miles from the mouth upstream to the deadline below Kalama Falls Hatchery and is generally seven days per week. Tributary harvest is managed to attain the Kalama Hatchery adult broodstock escapement goal.

Table 3.3.1.1: Kalama River Hatchery sub-yearling fall Chinook fishery contributions.

Brood Years: 2000-2009[b] Fishery Years:2004-2013			
	Average SAR%[a]	**0.26**	**0.27**
Agency	***Non-WA Fishery***	***% of total Survival***	
		Fallert Cr.	***KFH***
ADFG	All	*7.6*	*7.3*
CDFO	All	*22.2*	*24.1*
CDFW		*0.3*	----
NMFS	All	*1.8*	*0.7*
Agency	***OR Fishery***	***Fallert Cr.***	***KFH***
ODFW	10- Ocean Troll	*2.8*	*1.1*
ODFW	21- Columbia R. Gillnet	*2.8*	*4.1*
ODFW	40- Ocean Sport	----	*0.4*
ODFW	44- Columbia R. Sport	*1.6*	*2.9*
ODFW	45- Estuarine Sport (Buoy 10)	*1.3*	*2.1*
Agency	***WA Fishery***	***Fallert Cr.***	***KFH***
WDFW	10- Ocean Troll	*1.9*	*2.4*
MAKA	15- Treaty Troll	*0.5*	*0.7*
WDFW	15- Treaty Troll	*0.7*	*0.8*
WDFW	22- Coastal Gillnet	----	*0.2*
WDFW	23- Mixed Net and Seine	----	*0.1*
WDFW	41- Ocean Sport- Charter	*1.0*	*1.4*
WDFW	42- Ocean Sport- Private	*3.2*	*2.6*
WDFW	45- Estuarine Sport (Buoy 10)	----	*0.3*
WDFW	46- Freshwater Sport[c]	*3.5*	*3.5*
WDFW	50- Hatchery Escapement	*14.7*	*18.6*
WDFW	50- Hatchery Escapement (Strays)[d]	*1.0*	*2.9*

WDFW	54- Spawning Grounds[e]	*33.3*	*23.8*
	Total	***100.0***	***100.0***

Source: RMIS 2014.

[a] Average SAR% = (tags recovered/tags released).

[b] 2009 data is preliminary, and represents a minimum estimate.

[c] Freshwater Sport based on WDFW Catch Record Card (CRC) data.

[d] Includes recoveries at Cowlitz, Lewis, Marblemount and North Toutle Hatcheries.

[e] Includes recoveries in WRIA 26, 27 and 28.

3.4 Relationship to habitat protection and recovery strategies.

The following processes have included habitat identification problems, priority fixes and evolved as key components to *The Lower Columbia Salmon Recovery and Fish and Wildlife Sub-basin Plans* (Volume 1; Clark, Cowlitz, Lewis, Skamania and Wahkiakum Counties, December 15, 2004).

Sub-Basin Planning - Regional sub-basin planning processes include the *Kalama River Sub-basin Salmon and Steelhead Production Plan*, September 1, 1990 with a more recent Draft Kalama River Sub-basin Summary (May 17, 2002) was prepared for the *Northwest Power Planning Council* (NPPC). The sub-basin efforts provided initial building blocks for the LCFRB regional recovery plan. WDFW Region 5 staff is involved in fish and wildlife planning and technical assistance in concert through the LCFRB including the role of fish release programs originating from Kalama Complex. Staff is assessing the risks posed by the hatchery program using the *Benefit-Risk Assessment Procedure* (BRAP) in tandem with the LCFRB recovery plan.

Habitat Treatment and Protection - Ecosystem Diagnosis and Treatment (EDT) compares habitat today to that of the basin in a historically unmodified state. EDT has been modeled for productivity in the Kalama Basin in The Lower Columbia Salmon Recovery and Fish and Wildlife Sub-basin Plans. WDFW is also conducting a Salmon Steelhead Habitat Inventory Assessment Program (SSHIAP), which documents barriers to fish passage. WDFW's habitat program issues hydraulic permits for construction or modifications to streams and wetlands. This provides habitat protection to riparian areas and actual watercourses within the watershed.

Limiting Factors Analysis (LFA) - A WRIA 27 includes three major watersheds; the Kalama River, the Lewis River (North Fork), and the East Fork Lewis River. The LFA was conducted by the W*ashington State Conservation Commission* (January 2000). Loss of channel diversity, increased sedimentation, reduced stream flows, habitat constriction due to effects of irrigation withdrawn, water temperature, and inundation and loss of spawning/rearing habitat through dam construction, and fragmentation of habitat all affect productivity of natural salmonid populations within the watershed. The Lower Kalama River Hatchery presents a partial barrier to migration up Hatchery(Fallert) Creek during low flows. Reports for each WRIA are available at http://scc.wa.gov/directory/.

3.5 Ecological interactions.

(1) *Salmonid and non-salmonid fishes or species that could negatively impact the program:* Out-migrant hatchery fish can be preyed upon through the entire migration corridor from the river sub-basin to the mainstem Columbia River and estuary. Northern pikeminnows and introduced spiny rays, as well as avian predators, including gulls, mergansers, cormorants, belted kingfishers, great blue herons and night herons in the Columbia mainstem sloughs, can prey on steelhead smolts. Mammals that can take a heavy toll on migrating smolts and returning adults include: harbor seals, sea lions, river otters and orcas

(2) *Salmonid and non-salmonid fishes or species that could be negatively impacted by the program:* Co-occurring natural salmon and steelhead populations in local tributary areas and the Columbia River mainstem corridor areas could be negatively impacted by program fish. Of primary concern are the ESA listed endangered and threatened salmonids: Snake River

fall-run Chinook salmon ESU (threatened); Snake River spring/summer-run Chinook salmon ESU (threatened); Lower Columbia River Chinook salmon ESU (threatened); Upper Columbia River spring-run Chinook salmon ESU (endangered); Columbia River chum salmon ESU (threatened); Snake River sockeye salmon ESU (endangered); Upper Columbia River steelhead ESU (endangered); Snake River Basin steelhead ESU (threatened); Lower Columbia River steelhead ESU (threatened); Middle Columbia River steelhead ESU (threatened); and the Columbia River distinct population segment of bull trout (threatened). Listed fish can be impacted through a complex web of short and long term processes and over multiple time periods which makes evaluation of this a net effect difficult. WDFW is unaware of studies directly evaluating adverse ecological effects to listed salmon. In addition the program may have unknown impacts on eulachon populations in the basin.

(3) *Salmonid and non-salmonid fishes or other species that could positively impact the program.* Multiple programs including fall Chinook, coho and steelhead programs are released from the Kalama Hatchery and limited natural production of Chinook, coho, chum and steelhead occurs in this system along with non-salmonid fishes (sculpins, lampreys and sucker etc.).

(4) *Salmonid and non-salmonid fishes or species that could be positively impacted by the program.* Chinook smolts can be preyed upon release thru the entire migration corridor from the river sub-basin to the mainstem Columbia River and estuary. Northern pikeminnows and introduced spiny rays in the Columbia mainstem sloughs can prey on steelhead smolts as well as avian predators, including gulls, mergansers, cormorants, belted kingfishers, great blue herons and night herons. Mammals that benefit from migrating smolts and returning adults include: harbor seals, sea lions, river otters and orcas. Except for yearling coho and steelhead, these species may serve as prey items during the emigration through the basin. Hatchery fish provide an additional food source to natural predators that might otherwise consume listed fish and may overwhelm established predators providing a beneficial, protective effect to co-occurring wild fish. Hatchery releases can also behaviorally encourage mass emigration of multiple species through the watershed, reducing residency. Many watersheds in the Pacific Northwest appear to be nutrient-limited (Gregory et al. 1987; Kline et al. 1997) and salmonid carcasses can be an important source of marine derived nutrients (Levy 1997). Carcasses from returning adult salmonids have been found to elevate stream productivity through several pathways, including:

a) the releases of nutrients from decaying carcasses has been observed to stimulate primary productivity (Wipfli et al. 1998);
b) the decaying carcasses have been found to enrich the food base of aquatic invertebrates (Mathisen et al. 1988); and
c) Juvenile salmonids have been observed to feed directly on carcasses (Bilby et al. 1996).

4 SECTION 4. WATER SOURCE

4.1 Provide a quantitative and narrative description of the water source (spring, well, surface), water quality profile, and natural limitations to production attributable to the water source.

Table 4.1.1: Water sources for the Kalama Hatchery Complex.

Facility	Water Source	Water Right		Available Water Flow	Avg Water Temp. (°F)	Usage	Limitations
		Record/Cert. No	Permit No.				
Kalama Falls	Kalama River (surface) pump	S2-CV2P641/ VOL2P535	-----	265 cfs	43-51	Ladder and fishway; Adult holding,	Temps in lower river can reach the 70s in the summer

						incubation and rearing	
	Unnamed creek (surface) gravity intake	S2-*18989CWRIS/ 09625	14224	3 cfs		Incubation and rearing	No rearing during summer months due to temps and low water.
	Unnamed creek (surface) gravity intake	S2-*18990CWRIS/ 09624	14225	2 cfs			
Fallert Creek	Fallert Creek (surface) gravity intake	S2-21721C WRIS	-----	13 cfs	38-53	Adult holding, incubation and rearing	Limited water during summer months due to low flows. Temps in lower river can reach the 70s in the summer.
	Fallert Creek (surface) gravity intake	S2-25509C WRIS	-----	12 cfs	38-53		

Source: Phinney 2006, WDOE Water Resources Explorer 2014, WDFW Hatchery Data.
Note: S2-CV2P641 at Kalama Falls Hatchery covers a diversion of 8.75 cfs to Fallert Creek Hatchery.

Kalama Falls Hatchery. In the fall/winter of 2000/2001, a new intake pump station was constructed with FEMA monies after the 1996 flood damaged the facility. Five new pumps were installed capable of delivering approximately 16 cfs for rearing while two incubation pumps deliver 4 cfs. In 2010 a sixth pump rated at 3.2 cfs was added to the intake station increasing water pumping capacity to 19.2 cfs. A settling pond for incubation water was completed in 2002. Additionally, there are two surface water gravity intakes on unnamed creeks – one near the hatchery and one on the other side of the river – and because of steep gradients have been determined by WDFW to be non-fish bearing.

Water rights are formalized through the Washington Department of Ecology, and were obtained in 1953 and 1965.

Fallert Creek Hatchery. The intake structure is located near Fallert Creek, RM 0.5. Water can be gravity-fed from the creek intake providing up to 25cfs depending on weather and stream conditions. Pumps need to be used when dewatering becomes a concern late summer and early fall. The river intake is located adjacent to the hatchery with a four chambered pump system which can provide up to 8.7cfs (covered under S2-CV2P641 at Kalama Falls). Reuse water is available from the large earthen pond to the asphalt pond located near the Kalama River Road.

Water rights are formalized through the Washington Department of Ecology, and were obtained in 1973 and 1980.

NPDES Permits:

These facilities operate under the "*Upland Fin-Fish Hatching and Rearing*" *National Pollution Discharge Elimination System* (NPDES) general permit which conducts effluent monitoring and reporting and operates within the limitations established in its permit administered by the Washington Department of Ecology (DOE).

Discharges from the cleaning treatment system are monitored as follows:

- *Total Suspended Solids* (*TSS*) 1 to 2 times per month on composite effluent, maximum effluent and influent samples.
- *Settleable Solids* (*SS*) 1 to 2 times per week on effluent and influent samples.

- *In-hatchery Water Temperature* - daily maximum and minimum readings.

Table 4.1.2: Record of NPDES permit compliance.

Facility/ Permit #	Reports Submitted Y/N			Last Inspection Date	Violations Last 5	Corrective Actions Y/N	Meets Compliance Y/N
	Monthly	Qtrly	Annual				
Kalama Falls WAG13-1039	Y	Y	Y	5/2/2006	3	N	Y
Fallert Creek WAG13-1053	Y	Y	Y	5/2/2006	1	N	Y

Source: Ann West, WDFW Hatcheries Headquarters Database 2013.

Table 4.1.3: List of NPDES violations over the last five years (2008-2012).

Facility	Month/ Year	Parameter	Sample Type	Result/ Violation	Permit Limit	Comment	Action
KFH	Dec 2010	TSS	Avg Net Composite	6.87 mg/L	5.0 mg/L	High water event.	NA
	Jun 2011	TSS	Drawdown Max Grab	155.4 mg/L	100.0 mg/L	Late sampling and pond half cleaned.	Staff increased
	Feb 2012	TSS	Max Net Grab	23.8 mg/ L	15.0 mg/L	High river flows	NA
Fallert Cr	Aug 2010	TSS	Avg Net Composite	7.5 mg/L	5.0 mg/L	High river flow and heavy rains.	NA

Source: Ann West, WDFW Hatcheries Headquarters Database 2013.
Note: These violations did not result in non-compliance with NPDES permit.

4.2 Indicate risk aversion measures that will be applied to minimize the likelihood for the take of listed natural fish as a result of hatchery water withdrawal, screening, or effluent discharge.

Kalama Falls Hatchery. The intake was rebuilt in 2001, and is in compliance.

Fallert Creek. The intake screens are in compliance with state and federal guidelines (NMFS 1995, 1996), but do not meet the current *Anadromous Salmonid Passage Facility Design* criteria (NMFS 2011). WDFW is in the process of designing a new river intake system to meet NOAA-NMFS compliance (Mitchell Act Intake and Fish Passage Study Report 2003) and has included it in the 2013-2015 Capital Budget Request. A feasibility study was funded using Pacific Coast Salmon Recovery Funds in 2011 and completed in 2012.

5 SECTION 5. FACILITIES

5.1 Broodstock collection facilities (or methods).

Modrow Trap. The Modrow weir operates August through mid-October. A temporary rack is located on the Kalama River at RKm 4.8, a short distance below Modrow Bridge, and blocks upstream-migrant Chinook salmon, but currently has enough gap space to allow upstream and downstream migration of other species currently. Broodstock are collected at the trap located on the left bank and need to be trucked to Kalama Falls or Fallert Creek Hatcheries; transport time is around 20 and 10 minutes, respectively. Adults are transferred from the trap via overhead brail into a 1,500 gallon tanker truck and moved to either Kalama Falls Hatchery or Fallert Creek Hatchery.

The Modrow Trap has been used since 1968. WDFW proposed upgrading the site in 2011. Modifications include expanding the trap to increase holding capacity, concentrating flow to increase fish attraction, new crowder, and fish movers (pescalator and sorting tubes). The existing trap is 720 sq. ft.; upgrades would increase trap size to 1,295 sq. ft. Picket spacing will be reduced to 1.5" once the facility is complete fully functional. This is expected to take place in 2014.

Kalama Falls Hatchery. A trap at the Kalama Falls Hatchery operates 365 days a year. Fish volitionally enter the trap via a step and pool ladder at Kalama Falls Hatchery. Adults are transferred from the trap via overhead brail into a 1,500 gallon tanker truck, and hauled a short distance (150 m) to the 60'x10'x 5' concrete sorting pond (see HGMP section 5.3).

Fallert Creek Hatchery. The 55,000 cu-ft asphalt pond located near the Kalama River Road is converted to an adult holding pond with a step and pool ladder from August to mid-November. Dam boards are placed in the effluent structure of the pond to create the step and pool ladder. A temporary weir is placed in Fallert Creek immediately upstream of the entrance of the ladder during this timeframe.

5.2 Fish transportation equipment (description of pen, tank truck, or container used).

Table 5.2.1: Transportation equipment available at Kalama Hatchery Complex.

Equipment Type	Capacity (gallons)	Supp. Oxygen (y/n)	Temp. Control (y/n)	Norm. Transit Time (minutes)	Chemical(s) Used	Dosage (ppm)
Tanker Truck	1,500	Y	N	10-20	Sodium chloride (Salt)	5,000 ppm (~0.5%)

Adults are transported from the Modrow Trap to Kalama Falls and Fallert Creek hatcheries via 1500-gallon fiberglass tanker truck equipped with re-circulating pumps and supplemental oxygen system and adult release gate; transport time is around 10 minutes and 20 minutes to Fallert Creek and Kalama Falls hatcheries, respectively. Fish in surplus of broodstock needs may also be recycled downstream for additional harvest opportunities.

5.3 Broodstock holding and spawning facilities.

Table 5.3.1: Holding facilities available, Kalama Falls Hatchery.

Ponds (No.)	Pond Type	Volume (cu. ft.)	Length (ft.)	Width (ft.)	Depth (ft.)	Flow (gpm)	Max. Flow Index	Max. Density Index
1	Concrete adult sorting pond	3,000	60	10	5	500	1.61	0.20

Kalama Falls Hatchery. Fish in the 60'x10'x 5' concrete sorting pond are sorted three to five times per week; unmarked (natural-origin) fish are incorporated into broodstock for the integrated program. Fish kept for broodstock are moved to a 10,800 cu-ft holding pond with 500-800 gpm water flow (see also **Table 5.5.1**). Fish held for spawning are treated with formalin at 1:6,000 for fungus and parasite control. Early Chinook arrivals may be inoculated with oxytetracycline for *furunculosis* control at a rate of 0.5cc/10lbs of fish depending on water temperature. Water temperatures can range from a high of 67°F to a low of 46°F.

Fallert Creek Hatchery. Broodstock have been spawned at Fallert Creek since 2009. Adults are seined, sorted, killed and spawned directly from the 55,000 cu-ft asphalt adult holding pond, supplied with water at 1,200 gpm (see HGMP section 5.1). Fish not ready to spawn are returned to the pond for further maturation. Spawning for this program takes place in a covered area.

5.4 Incubation facilities.

Table 5.4.1: Incubation vessels available at Kalama Hatchery Complex.

Facility	Type	Units (number)	Size	Flow (gpm)	Volume (cu. ft.)	Loading (eggs/unit)
KFH	Heath Vertical Stack Tray Units (14 trays/stack)	84 1,176 trays	24" x 25' 'x 4"	5	0.55/tray	8,000
	Free Style eyeing unit	15	41" x 15" x 21"	20	7.48/unit	300,000
Fallert Creek	Vertical Stack Tray Units (14 trays/stack)	28	24" x 25' 'x 4"	5	0.55/tray	8,000

	Deep Troughs	4	16’ x 2.8’ x 24”	15	91	
	Shallow Troughs	16	15’ x 1’ x 8”	n/a	10	n/a
	Freestyle eyeing unit	3	41”x15”x21”	20	7.48/unit	300,000

Combined lots of approximately 300,000 eggs are loaded into free-style eyeing units for or in vertical stack trays at 8,000/tray. Eggs are treated with iodophor. Kalama River water is used for most of the incubation; flow starts at 5 gpm per stack, DO averages 10.7 ppm.

5.5 Rearing facilities.

Table 5.5.1: Rearing facilities available, Kalama Falls Hatchery.

Ponds (No.)	Pond Type	Volume (cu. ft.)	Length (ft.)	Width (ft.)	Depth (ft.)	Flow (gpm)	Max. Flow Index	Max. Density Index
12	Standard Concrete Raceways	4,800	80	20	3	600	1.61	0.20
5	Rearing/Adults Ponds	10,800	60	40	5	800	1.61	0.20
1	Rearing/Adults Pond	9,000	60	30	5	800	1.61	0.20
4	Fiberglass Intermediate Deep Troughs	91	16	2.8	2	20	1.61	0.20

Kalama Falls Hatchery. Fall Chinook are typically reared in eight raceways and all of the rearing/adult ponds.

Table 5.5.2: Rearing facilities available, Fallert Creek Hatchery.

Ponds (No.)	Pond Type	Volume (cu. ft.)	Length (ft.)	Width (ft.)	Depth (ft.)	Flow (gpm)	Max. Flow Index	Max. Density Index
8	Concrete Raceways	6,400	80	20	4	600	1.61	0.20
1	Asphalt Pond - (adult holding or juvenile rearing)	55,000	190	55	5.3	1,200	2.42	0.05
1	Gravel-bottom pond	55,000	100	90	5.3	1,500	2.42	0.05
9	Fiberglass Intermediate Deep Troughs	91	16	2.8	2	20	1.61	0.20

Fallert Creek Hatchery. Fall Chinook are typically reared in five raceways and both large rearing ponds.

5.6 Acclimation/release facilities.

Fish are released from the ponds at their respective facilities (see HGMP section 5.5) directly to the river. Smolts from Kalama Falls Hatchery are force-released; releases from Fallert Creek are volitional.

5.7 Describe operational difficulties or disasters that led to significant fish mortality.

In some years, low flows and high water temperatures lead to significant low dissolved oxygen levels. In 2003, staff cut back feeding regimes and released the program at Fallert Creek earlier, before program size was met (96-102 fpp).

Flooding and associated debris and sediments chronically affect fish production at Kalama Falls Hatchery.

5.8 Indicate available back-up systems, and risk aversion measures that will be applied, that minimize the likelihood for the take of listed natural fish that may result from equipment failure, water loss, flooding, disease transmission, or other events that could lead to injury or mortality.

- All pumps, broodstock holding, incubation and rearing receptacles have water loss alarms.
- Staff is available 24/7 to respond to pump failure, water loss, and flooding events.
- Aeration pumps are used to maximize the water conditions in the adult collection pond during periods of low water quality which benefits fish held until sorting can be accomplished.
- Fish health protocols through broodstock collection, incubation and rearing phases are followed and monitored monthly.
- Broodstock collection is checked daily for program and listed fish.
- Staff monitors the trap operation daily to keep the numbers of fish stacking in the trap area to manageable volumes. Heavy volumes can create density problems for listed fish if they are not removed expeditiously.

6 SECTION 6. BROODSTOCK ORIGIN AND IDENTITY

Describe the origin and identity of broodstock used in the program, its ESA-listing status, annual collection goals, and relationship to wild fish of the same species/population.

6.1 Source.

Broodstock for the program are collected at the traps from adult recruits returning to the Kalama River. Broodstock for both facilities are collected at the Modrow Trap, a temporary rack upstream of tidewater, just below Modrow Bridge. Broodstock can also be collected at the Kalama Falls and Fallert Creek Hatcheries. Annual estimates of fall Chinook escapement will be made for the Kalama River up to Kalama Falls Hatchery.

6.2 Supporting information.

6.2.1 History.

Plants to the Kalama River began when Fallert Creek was completed in 1895; Kalama Falls Hatchery was completed in 1959. Fall Chinook are native to the Kalama River and were historically abundant. This is a mixed stock with composite production and is similar in life history to other fall Tule Chinook (SaSI 2002). It is probable that a significant number of natural spawners are hatchery strays and strays from other hatcheries within this GDU are common. Broodstock for both facilities are taken from the seasonally-operated Modrow Trap. Broodstock can also be collected at the Kalama Falls and Fallert Creek hatchery traps; broodstock have been collected at Fallert Creek Hatchery since 2009. There have been relatively few introductions of out-of-basin Chinook into the hatchery program (Myers et al. 2006).

6.2.2 Annual size.

Up to 1,445 adult pairs, not including jacks, are needed to achieve the established egg-take goal of 7.7-million (FBD 2014). This is based on an average fecundity of around 5,400 eggs/female, and a pre-spawning mortality of 5%.

6.2.3 Past and proposed level of natural fish in broodstock.

The level of natural origin fish in the returning broodstock was unknown prior to the start of mass-marking in 2005. WDFW moved to an integrated program, incorporating natural fish into the broodstock in 2010, the first year of mass-marked returns for all age classes. Improvement

will be completed on the Modrow Weir in 2014 to allow for better control of hatchery fish on the spawning grounds. See **Table 7.4.2.1** for last twelve years of broodstock collection.

Table 6.2.3.1: Integration rates for Kalama Falls Hatchery fall Chinook program, 2010-2013.

Year	pNOB	pHOS	PNI
2010	0.04	0.88	0.04
2011	0.14	0.93	0.13
2012	0.05	0.93	0.05
2013	0.16	0.91	0.15
Avg	**0.10**	**0.91**	**0.10**

Source: WDFW Hatchery Evaluation and Assessment Team Broodstock Tracking tables 2014.
Note: First year of mass-marked returns for all age classes was 2010.

6.2.4 Genetic or ecological differences.

The broodstock is derived from stock returning to the sub-basin. Straying of Lower River Hatchery (LRH) fall Chinook from a number of Oregon and Washington hatcheries is not unusual, and contributes to natural production. The overall result of straying and transfers of fall Chinook at lower Columbia River hatcheries is the development of a widely-distributed, blended hatchery stock. There are no known genotypic, phenotypic, or behavioral differences between hatchery or natural stocks in the sub-basin. Fall Chinook propagated through the program represent the indigenous lower Columbia stock. During years where insufficient numbers of adults return, eggs may be obtained from other lower Columbia River hatchery facilities with Tule Chinook, if available. No genetic analysis has been done on naturally spawning Kalama fall Chinook. Allozyme analysis of Kalama Hatchery fall Chinook sampled in 1988 and 1989 showed that they were genetically distinct from most other lower Columbia Tule fall Chinook but not significantly different from Abernathy Creek (Myers et al. 2006, in SaSI 2002).

6.2.5 Reasons for choosing.

The stock has a run-entry pattern and timing that provides harvest opportunities for fisheries in the sub-basin, lower Columbia mainstem/tributaries, and the Washington/Oregon Coast, and represent the indigenous lower Columbia stock. The broodstock chosen has the desired life history traits to meet these harvest goals (e.g. run-timing) that provides significant harvest to the ocean and lower Columbia River fisheries (e.g., Buoy 10).

6.3 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic or ecological effects to listed natural fish that may occur as a result of broodstock selection practices.

- Natural spawners will be integrated into the broodstock to represent the natural fall Chinook run throughout the season.
- Hatchery program fish are mass-marked.
- There are no known genotypic, phenotypic, or behavioral differences between either the hatchery stock or natural stock in the sub-basin.
- Holding pond procedures follow IHOT guidelines.
- Other listed fish encountered during the broodstock collection process will be returned directly to the river or passed into the upper watershed, with minimal handling and holding time.
- Any observed mortalities will be reported in the WDFW Hatcheries Headquarters Database.

7 SECTION 7. BROODSTOCK COLLECTION

7.1 Life-history stage to be collected (adults, eggs, or juveniles).

Adults returning to the Kalama River.

7.2 Collection or sampling design.

Modrow Trap. Broodstock are collected at the Modrow Trap, located on the Kalama River at RKm 4.8. The trap consists of a temporary rack-picket structure with a "V"-trap for capturing/holding adults. The trap structure is operated in the river during from August 1 through mid-October period. Captured adults are sorted and transferred to a tanker truck via a brail-hoist system and transported to Kalama Falls or Fallert Creek Hatcheries. The rack and picket structure is currently designed to allow upstream passage of steelhead, coho and cutthroat, and downstream migrant smolts. Fish are removed weekly, or as needed with heavy volumes to reflect run timing. The redesigned facility will have 1.5" picket spacing and will have fish handling capabilities for all species and return tubes to return no-target species back to the river.

Fish released upstream from Modrow arrive at the upper Kalama Falls trap approximately one week later, during the peak period. Egg-take goals are separated by an early component taken in late-September/early-October, and later component taken in late-October. Surplus fish can be used for nutrient enhancement in the sub-basin or donated to food banks.

Kalama Falls Hatchery. The trap at Kalama Falls operates 365 days a year, due to returns of spring and fall Chinook, Type-S and Type-N coho, and winter and summer steelhead. Fish volitionally enter the trap via a step and pool ladder at Kalama Falls Hatchery. Adults are transferred from the trap via overhead brail into a 1,500-gallon tanker truck, and moved to the sorting pond.

During the fall Chinook season, excess fish can be right opercle punched and recycled downstream. A small number of fish make the trip back to the Kalama Falls trap a second time; some are caught and others are assumed to spawn naturally. A detailed report on the distribution of these fish is not available. All fish returning to the hatchery are examined for tags and marks (including fin-clips and opercle punches). Information is reported to the WDFW Hatcheries Headquarters Database.

Table 7.2.1: Number of fall Chinook recycled for additional harvest opportunity returning to the Kalama Falls Hatchery trap.

Year	Total Return	Recycled Downstream	Returned to KFH
2008	4,255	274	15
2009	4,302	241	18
2010	3,129	328	34
2011	5,525	1,871	119
2012	5,285	790	27
2013	5,667	244	5

Source: WDFW hatchery records 2014.

Fallert Creek Hatchery. Broodstock have been spawned at Fallert Creek since 2009. The large asphalt pond located near the Kalama River Road is converted to an adult holding pond with a step and pool ladder from August to mid-November. Dam boards are placed in the effluent structure of the pond to create the step and pool ladder. A temporary weir is placed in Fallert Creek immediately upstream of the entrance of the ladder during this timeframe. Fish volitionally enter the pond via the temporary ladder.

Broodstock are also transferred from the Modrow Trap.

7.3 Identity.

Fall Chinook produced from this facility have been released mass-marked with an adipose fin-clip (AD) since brood year 2005, return year 2010. Prior to this, natural-origin fish could not be differentiated from hatchery-origin fish. A total of 250,000 sub-yearlings are also released AD+coded-wire tagged (CWT).

7.4 Proposed number to be collected:

7.4.1 Program goal (assuming 1:1 sex ratio for adults):

See HGMP section 6.2.2.

7.4.2 Broodstock collection levels for the last twelve years (e.g. 2002-13), or for most recent years available:

Table 7.4.2.1: Sex, composition and origin of Fall Chinook broodstock collected at Modrow Trap, and Kalama Falls and Fallert Creek Hatcheries 2002-2013.

Brood Year	Hatchery-origin			Natural-origin*		
	Females	Males	Jacks	Females	Males	Jacks
2002	955	1038	8			
2003	930	958	0			
2004	1,117	1,126	9			
2005	963	963	0			
2006	961	1030	20			
2007	836	988	19			
2008	1,192	1,272	13			
2009	1,478	1,595	24			
2010*	1,165	1,217	36	48	56	0
2011	1,233	1,254	29	176	203	6
2012	1,829	1,788	32	57	135	0
2013	1,301	1,177	44	173	283	8

Source: WDFW Hatchery Headquarters Database 2014.
Note: Broodstock have been collected at both Kalama Falls and Fallert Creek Hatcheries as of 2009.
*Returns in 2010 represent the first year all age classes were mass marked, in prior years it was not possible to determine the number of natural-origin fish returning.

7.5 Disposition of hatchery-origin fish collected in surplus of broodstock needs.

Fish at both facilities in surplus of broodstock and nutrient enhancement needs may be recycled downstream for additional sport harvest opportunity. No fall Chinook are passed above the Kalama Falls Hatchery. In high return years, fish may be donated to food banks.

7.6 Fish transportation and holding methods.

Adult fish are trucked via a tanker mounted on a flatbed truck from the Modrow Trap and the Kalama Falls trap to holding and sorting ponds (see HGMP section 5.2). Normal transport time from Modrow is 20 minutes to Kalama Falls, and 10 minutes to Fallert Creek Hatcheries.

Kalama Falls Hatchery. Fish kept for broodstock held in a 10,800 cu. ft. holding pond with 600-800 gpm water flow (see HGMP section 5.3).

Fallert Creek Hatchery. Fish kept for broodstock held in a 55,000 cu. ft. asphalt adult holding pond (see HGMP section 5.3).

7.7 Describe fish health maintenance and sanitation procedures applied.

Fish held for spawning are treated with formalin at 1:6000 for fungus and parasite control. Early Chinook arrivals may be inoculated with oxytetracycline for furunculosis control at a rate of 0.5cc/10lbs of fish.

The adult holding area is separated from all other hatchery operations. Disinfection procedures that prevent pathogen transmission between stocks of fish are implemented during spawning. Spawning implements are rinsed with an iodophor solution, and spawning area and implements are disinfected with iodophor solution at the end the spawning day.

7.8 Disposition of carcasses.

Carcasses can be used for nutrient enhancement, or disposed of at an approved upland site (landfill).

7.9 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic or ecological effects to listed natural fish resulting from the broodstock collection program.

- Every effort shall be made to promote local adaptation of this fall Chinook population.
- Limit out-of-basin transfers of eggs or fish, except in extreme cases.
- Fall Chinook have been mass-marked for Chinook programs since brood year 2005 (full adult returns for all age classes in 2010).
- Minimum of 500 adults collected.
- There are no known genotypic, phenotypic, or behavioral differences between either the hatchery stock or natural stock in the sub-basin.
- Holding pond procedures follow IHOT guidelines.
- Other listed fish will be released immediately, if encountered, during the broodstock collection process.

8 SECTION 8. MATING

Describe fish mating procedures that will be used, including those applied to meet performance indicators identified previously.

8.1 Selection method.

Cohorts are utilized as fish ripen over the entire run cycle, based on the historical run curve, to ensure that the run timing for the stock is maintained; males and females available on a given day are mated randomly. Spawning is conducted weekly, with the peak in mid-October. The spawning protocol mandates the use of a spawning population of at least 500 adults.

8.2 Males.

A ratio of 1:1 males to females is used. Jacks (2-year olds) can be incorporated into the broodstock as males up to 2% of the total number of spawned fish.

8.3 Fertilization.

Over all ratio of 1:1 is applied. For daily egg takes over 500,000, eggs from five females are spawned into a bucket (ovarian fluid is not drained), and the sperm from five males is then combined with the eggs. Egg takes at <500,000 per day utilize a true one to one spawning ratio.

Water hardening procedures with iodophor are followed after twenty minutes. Iodophor solution is used as rinse that is applied to hands and spawning implements per spawning. Iodophor foot baths are located at entrance to incubation room. Generally, sixty ovarian fluid and kidney/spleen samples are collected from female spawners to test for the presence of viral pathogens. Unmarked fish not used for integration needs are released downstream of the hatchery.

8.4 Cryopreserved gametes.

Cryopreserved gametes are not used.

8.5 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic or ecological effects to listed natural fish resulting from the mating scheme.

- Limit/eliminate out-of-basin transfers.
- Protocols for population size, fish health disinfection and genetic guidelines are followed.
- Spawn all collected mature broodstock if possible without regard to age, size, color or other physical characteristics. If not spawning all collected mature adults over the season, apply the same rationale to individual spawn days.
- Randomize mating and avoid selectivity beyond ripeness on a given spawn day.
- Use one male to one female as much as possible in order to ensure an equal genetic contribution.
- Do not re-use males except as part of specific spawning protocols. A given male should be used as the first mate for only one female total.

9 SECTION 9. INCUBATION AND REARING -Specify any management *goals* (e.g. "egg to smolt survival") that the hatchery is currently operating under for the hatchery stock in the appropriate sections below. Provide data on the success of meeting the desired hatchery goals.

9.1 Incubation:

9.1.1 Number of eggs taken and survival rates to eye-up and/or ponding.

Table 9.1.1: Total Egg Take for the Kalama Fall Chinook Program 2002-2013.

Brood Year	Egg-Take
2002	5,676,255
2003	5,262,005
2004	5,674,450
2005	5,146,106
2006	5,116,053
2007	4,592,019
2008	6,270,061
2009	7,825,985
2010	6,499,386
2011	7,698,836
2012	9,603,334
2013	7,678,458

Table 9.1.2: Survival rates (%) from egg-take to ponding, Kalama River fall Chinook.

Brood Year	Green-to-Eyed		Eyed Egg-to-Ponding	
	KFH	Fallert Cr*	KFH	Fallert Cr
2002	n/a	n/a	94.5	95.8
2003	95.8	n/a	97.7	97.7
2004	94.5	n/a	97.6	97.3
2005	95.1	n/a	97.8	98.0
2006	93.5	n/a	96.6	96.9
2007	95.7	94.7	97.6	99.0
2008	92.4	90.8	97.6	99.5
2009	93.9	92.0	98.0	99.5
2010	98.7	98.1	98.2	97.4
2011	89.9	99.4	96.8	93.1
2012	88.2	95.6	99.0	98.5

Source: WDFW hatchery records.
* Green eggs were not collected at Fallert Creek Hatchery prior to 2007.

9.1.2 Cause for, and disposition of surplus egg takes.

The program broodstock collection goals set in the annual Future Brood Document. Egg-takes are managed according to data/information of historical egg-takes at the facility, and are maintained within the ±5% guideline of the Section 7 permit. Viral sampling (60 fish lots) are conducted over the course of the season.

In the event that egg survival is higher than expected, WDFW Regional Managers will be contacted for instructions for disposition of the surplus in accordance with Regional policy and guidelines set forth in management plans/agreements and ESA permits.

9.1.3 Loading densities applied during incubation.

Eggs taken at Kalama Falls Hatchery are incubated in free-styles or vertical trays to the eyed-stage, then approximately 50% are transferred to Fallert Creek Hatchery. Eggs that are taken from broodstock at Fallert are incubated and reared on-station.

Eggs are placed in the stack incubators at 8,000 eggs/tray for eyeing and hatching. Removal of dead eggs, accurate enumeration and loadings are adjusted during this time. WDFW follows *Integrated Hatchery Operations Team* (IHOT) species-specific incubation recommendations for water quality, flows, temperature, substrate, and incubator capacities.

9.1.4 Incubation conditions.

IHOT species-specific incubation recommendations are followed for water quality, flows, temperature, substrate and incubator capacities. Incubation water temperature is monitored by thermograph and recorded and temperature units (TU) are tracked for embryonic development. Harmful silt and sediment is cleaned from incubation systems regularly while eggs are monitored to determine fertilization and mortality.

Combined lots of approximately 300,000 eggs are loaded into free-style eyeing units or in vertical stack trays at 8,000 eggs /tray. Eggs are treated with iodophor. Kalama River water is used for most of the incubation.

Eyed-eggs are treated with iodophor and formalin until eggs are ready to be shocked and picked. Egg-takes are divided between Kalama Falls and Fallert with each facility rearing approximately 50% of the program. Eyed-eggs are loaded into the stack incubators at 8,000 eggs/tray and incubated on surface water at a flow of 5 gpm. Dissolved oxygen (DO) content is monitored and

are at a minimum criteria of 8 ppm ranging up to ambient saturation levels; aerators could be employed if DO is below target. Siltation is controlled with rodding as needed.

9.1.5 Ponding.

Fry are typically ponded to the raceways starting in late-January when the yolk slit is closed to approximately 1-mm wide (approximately 1,600 TUs) or KD factor (95% yolk absorption).

9.1.6 Fish health maintenance and monitoring.

Staff conducts daily inspection visual monitoring and sampling from eye, fry, fingerling and sub-yearling stages. As soon as potential problems are seen these concerns are immediately communicated to the WDFW fish health specialist. In addition fish health specialists conduct inspections monthly. Potential problems are managed promptly to limit mortality and reduce possible disease transmission. Disease treatment varies with the pathogen encountered but is generally antibiotic in nature for bacterial infections and bath or drip treatments with chemotheraputants for external infections.

See also **Attachments 1** and **2** for fish health and virology information.

9.1.7 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish during incubation.

- Limit/eliminate out-of-basin transfers.
- IHOT and WDFW fish health guidelines followed.
- Multiple units are used in incubators.
- Splash curtains can isolate incubators.
- Temperature, dissolved oxygen, and flow are monitored.
- Dead eggs are discarded in a manner that prevents disease transmission.

9.2 Rearing:

9.2.1 Provide survival rate data (*average program performance*) by hatchery life stage (fry to fingerling; fingerling to smolt) for the most recent twelve years (2002-12), or for years dependable data are available.

Table 9.2.1.1: Survival rates (%) of fall Chinook from ponding to release, at Kalama River and Fallert Creek Hatcheries.

Brood Year	Survival Rates (%) Ponding to Release	
	KFH	Fallert Cr
2002	94.1	95.6
2003	96.3	96.3
2004	95.7	96.1
2005	85.6	96.0
2006	93.5	98.1
2007	99.2	98.5
2008	99.0	101.2
2009	93.8	99.1
2010	94.1	97.4
2011	92.2	99.2
2012	96.3	94.9

Source: WDFW Hatchery Records 2014.

9.2.2 Density and loading criteria (goals and actual levels).

Loading and density levels at WDFW hatcheries conform to standards and guidelines set forth in *Fish Hatchery Management* (Piper et. al. 1982), the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006). IHOT standards are followed for water quality, alarm systems, predator control measures to provide the necessary security for the cultured stock, loading and density.

Densities are generally kept at or below 3.3 lbs /gpm and 0.5 lbs /cu ft. with some rearing vessels reaching 5.2 lbs/gpm and 0.65lbs/cu ft. before release. Tank and raceway maximum loading for early rearing is 132 lbs for the tanks at 40 gpm (3.3 lbs/gpm) and 800 lbs per raceway at 300 gpm.(2.66 lbs/gpm). The final loading per raceway is approximately 3200 lbs. at 700 gpm (10.6 lbs/gpm).

9.2.3 Fish rearing conditions

Table 9.2.3.1: Monthly average surface water temperature (°F), Kalama River.

Month	Average Water Temperature (ºF)
January	42
February	43
March	46
April	48
May	51
June	55
July	61
August	60
September	56
October	49
November	47
December	43

Source: WDFW Hatchery Records 2014.

IHOT standards are followed for water quality, alarm systems, predator control measures (netting), loading and density.

Fish are reared on water from Kalama River, the unnamed creek, and reuse water (final rearing). Temperature, dissolved oxygen (DO) and pond turn-over rate are monitored and recorded daily; water temperatures during the rearing cycle range from 32° to 61°F (Table 9.2.2), DO averages around 10.7 ppm in March/April. In the event that both facilities are at capacity when there are low dissolved oxygen (DO) levels, fish would be released early after consulting with Regional management. Otherwise, fish would be split to reduce densities.

Settleable solids, unused feed and feces are removed regularly to ensure proper cleanliness of rearing containers. All ponds are vacuumed as needed. Predator netting over the raceways minimizes predation. The ponds are pressure-washed between broods. All raceways and rearing ponds can be utilized to facilitate mass-marking of the program.

Fish are mass-marked when they reach about 250 fpp. This generally occurs in mid-April, and can continue through June.

9.2.4 Indicate biweekly or monthly fish growth information (*average program performance*), including length, weight, and condition factor data collected during rearing, if available.

Table 9.2.4.1: Monthly fish growth information by length (mm), weight (fpp), condition factor and growth rate.

Rearing Period	Length (mm)	Weight (fpp)	Growth Rate
February	36.7	1,101	NA
March	52.4	378	0.657
April	70.4	156	0.587
May	87.8	80	0.487
June	87.8	80	NA
July	87.8	80	NA

Source: WDFW Hatchery Records 2014.

The first egg takes are ponded in February and typically reach release size (80 fpp) in late-May/early-June, depending on water temperatures and rearing conditions. Fish are released in June and July when the average size of fish in a given rearing container reaches 80 fpp. Later egg-takes may be released as late as mid-July.

9.2.5 Indicate monthly fish growth rate and energy reserve data (average program performance), if available.

See HGMP section 9.2.4. No energy reserve data available.

9.2.6 Indicate food type used, daily application schedule, feeding rate range (e.g. % B.W./day and lbs/gpm inflow), and estimates of total food conversion efficiency during rearing (*average program performance*).

Fish are given variety of diet formulations including starter, crumbles and pellets; the food brand used may vary, depending on cost and vendor contracts. Feeding frequencies varies depending on the fish size and water temperature, and usually begin at 7-8 feedings/7 days a week, and end at 1-4 feedings/week. Fall Chinook are typically fed around 2.5% B.W./day, depending upon water temperature and weather conditions. Feed rate is applied in accordance with program goals not to exceed 0.1-0.15 pounds feed per gallon in-flow, depending on fish size. Average season conversion rates generally are no greater than 1.3:1.0.

9.2.7 Fish health monitoring, disease treatment, and sanitation procedures.

Monitoring. Policy guidance includes: *Fish Health Policy in the Columbia Basin*. Details hatchery practices and operations designed to stop the introduction and/or spread of any diseases within the Columbia Basin. Also, *Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries* (Genetic Policy Chapter 5, IHOT 1995). A fish health specialist inspects fish monthly and checks both healthy and presence of symptomatic fish. Based on pathological or visual signs by the crew, age of fish and the history of the facility, the pathologist determines the appropriate tests. External signs such as lesions, discolorations, and fungal growths will lead to internal examinations of skin, gills and organs. Kidney and spleen are checked for BKD. Blood is checked for signs of anemia or other pathogens. Additional tests for virus or parasites are done if warranted (see **Attachment 1** - Fish Health Monitoring history and **Attachment 2** for Virology Sampling reports).

Disease Treatment. As needed, appropriate therapeutic treatment will be prescribed to control and prevent further outbreaks. At Fallert Creek, fish may be treated with formalin for *Costia* and florfenicol for furunculosis. Sub-yearlings at Kalama Falls Hatchery may be treated with Parasite-S® (formalin) for *Ichthyophthirius* and fungus control in adults. Oxytetracycline is used to treat

furunculosis. Mortality is collected and disposed of at a landfill. Fish health and/or treatment reports are kept on file (see also **Attachment 1**: Fish Health Monitoring summaries).

Sanitation. All eggs brought to the facility are surface-disinfected with iodophor (as per disease policy). Every effort is made to prevent the horizontal spread of pathogens by splashing water. All equipment (nets, tanks, boots, etc.) is disinfected with iodophor between different fish/egg lots. Different fish/egg lots are physically isolated from each other by separate ponds or incubation units. Footbaths containing disinfectant are strategically located on the hatchery grounds to prevent spread of pathogens. Mortalities are collected and disposed of at a landfill. Fish Health and/or treatment reports are kept on file (see **Attachment 1** for Fish Health monitoring history).

9.2.8 Smolt development indices (e.g. gill ATPase activity), if applicable.

Gill ATPase activity is not measured. Fish size at release time is critical to the readiness for migration. The migratory state of the release population is determined by fish behavior. Aggressive screen and intake crowding, swarming against sloped pond sides, a leaner (0.80 – 0.90) condition factor (K), a silvery physical appearance and loose scales during feeding events are signs of smolt development.

9.2.9 Indicate the use of "natural" rearing methods as applied in the program.

Not applicable.

9.2.10 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish under propagation.

See HGMP sections 5.8, 6.3, 7.9 and 9.1.7.

10 SECTION 10. RELEASE

Describe fish release levels, and release practices applied through the hatchery program.

10.1 Proposed fish release levels.

Table 10.1.1: Proposed release levels (maximum number).

Age Class	Maximum Number	Size (fpp)	Release Date	Location
Sub-yearling	7,000,000	80.0	May/July	Kalama River

Source: WDFW Future Brood Document 2014.
Note: 80 fpp = 88 mm fork length (fl).

10.2 Specific location(s) of proposed release(s).

Stream, river, or watercourse:	Kalama R (WRIA 27.0002)	Fallert Creek (WRIA 27.0017)
Release point:	RKm 16.1	RKm 8.2; trib to Kalama at RKm 4.9
Major watershed:	Kalama Sub-Basin	
Basin or Region:	Lower Columbia River	

10.3 Actual numbers and sizes of fish released by age class through the program.

Table 10.3.1: Number, average size (fpp), CV and date of release, Kalama River sub-yearling Chinook, by facility and year.

Release Year	Kalama Falls Hatchery				Fallert Creek Hatchery			
	Number	Avg Size (fpp)	CV	Date	Number	Avg Size (fpp)	CV	Date
2002	2,565,346	73.8	6.88	6/18, 21	2,433,562	72.0	6.45	6/6-5, 10-15, 24
2003	2,545,640	77.2	8.28	6/5,7,9,15	2,381,626	97.6	6.28	6/8, 18, 19
2004	2,503,175	74.3	7.54	6/14, 21	2,225,639	77.8	7.48	6/14-21
2005	3,141,233	78.2	7.95	6/5	2,065,759	79.0	7.94	6/27
2006	2,012,085	71.1	7.95	6/13,22,28; 7/11	2,016,536	81.3	8.72	6/13-30
2007	2,374,761	72.5	6.56	6/21, 28	2,144,016	86.3	7.85	6/6-30; 7/2
2008	2,142,109	79.3	4.42	6/25; 7/1,7	1,750,565	79.2	6.18	6/22-7/6
2009	2,957,203	75.5	4.35	6/19, 24, 29; 7/6, 8	2,569,453	79.2	7.91	6/4-7, 17-20; 7/4
2010	3,863,814	78.0	5.18	6/15, 23, 28; 7/7, 13, 20	3,419,678	80.0	6.40	6/14-7/2
2011	3,292,823	75.0	6.80	6/3, 15, 22, 30; 7/5	2,634,799	76.3	n/a	6/13-17; 7/1-4
2012	3,193,343	77.3	6.97	6/20, 26-27; 7/11	3,545,564	78.6	6.67	6/8-15, 6/23-7/2, 7/6
2013	3,157,011	78.2	6.71	6/11, 18, 25	3,522,335	79.9	n/a	6/1-4, 10

Source: WDFW Hatcheries Headquarters Database 2014.
Note: 70.0 fpp = 92 mm fork length (fl); 80.0 fpp = 88 mm fl; 90 fpp = 84 mm fl.

10.4 Actual dates of release and description of release protocols.

Sub-yearling Chinook are generally released in mid-June/early-July; however, due to the size of the program and possible issues with low water and high temperatures, fish may be released in May. See **Table 10.3.1** for actual release dates. The release period of program fish lies within the natural out migration time frame of naturally-produced Tule fall Chinook. River temperature and discharge can also be determinants of the date of release. Fish in the ponds can be drawn down slowly and volitionally released over several days, after which remaining fish are forced out. Fish in the raceways are force-released.

Kalama Falls Hatchery. Chinook releases start in early- to mid-June, and end in mid-July. Fish are force-released.

Fallert Creek Hatchery. Chinook releases start in early- to mid-June, and end in early July.

10.5 Fish transportation procedures, if applicable.

Some juvenile fish may be transported between facilities in order to facilitate mass marking. Fish are moved via a fish pump and 1,500 gallon tanker truck (see HGMP section 5.2).

10.6 Acclimation procedures (methods applied and length of time).

Fish for this program are reared, acclimated, and released directly from the rearing ponds at Kalama Falls and Fallert Creek Hatcheries. All production is reared with surface and re-use water. All fish are programmed to be in a smolt size before release; releases fall within the normal migration time of natural fish.

10.7 Marks applied, and proportions of the total hatchery population marked, to identify hatchery adults.

Table 10.7.1: Marks applied, by brood year, age class and mark-type.

Brood Year	Age Class	Number	Mark-Type
2014	**Sub-yearlings**	250,000	AD+CWT
		6,750,000	AD-only

Source: WDFW Future Brood Document 2014.

Fish have been 100% mass-marked (adipose fin-clipped) since 2005. In addition, 125,000 sub-yearlings are released AD + coded-wire tag (AD+CWT) from each facility to help determine origin and straying rates.

Scale samples are read at WDFW Headquarters Olympia to determine age composition for run re-construction. Snouts collected from the adipose fin-clipped adults are dissected, recovered and read at the WDFW CWT Lab in Olympia. CWT data is reported annual to RMIS.

10.8 Disposition plans for fish identified at the time of release as surplus to programmed or approved levels.

The program guidelines for annual broodstock/egg-take collection are managed to prevent any surpluses, and maintained within the ±5% guideline. In the event of surplus >10%, WDFW Regional Managers will in accordance with regional policy and guidelines set forth in management plans/agreements and ESA permits, and after consultation with NMFS, instruct hatchery staff for disposition of the surplus.

10.9 Fish health certification procedures applied pre-release.

All fish are examined for the presence of "reportable pathogens" as defined in the P*acific Northwest Fish Health Protection Committee* (PNFHPC) disease control guidelines, within three weeks prior to release. Fish transfers into the sub-basin are inspected and accompanied by notifications as described in IHOT and PNFHPC guidelines.

Prior to release, the population health and condition is established by the Area Fish Health Specialist. This is commonly done 1-3 weeks pre-release and up to six weeks on systems with pathogen-free water and little or no history of disease. Prior to this examination, whenever abnormal behavior or mortality is observed, staff also contacts the Area Fish Health Specialist. The fish specialist examines affected fish, and recommends the appropriate treatment. Reporting and control of selected fish pathogens are done in accordance with the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006) and IHOT guidelines.

10.10 Emergency release procedures in response to flooding or water system failure.

In the event of a water system failure, screens would be pulled to allow fish to exit the ponds or in some cases they can be transferred into other rearing vessels to prevent an emergency release. WDFW also has emergency response procedures for providing back-up pumps, transport trucks, etc. in cases of emergency. In cases of severe flooding the screens are not pulled because flood waters rise to the point where they breach the ponds. Past experience has shown that the fish tend to lie on the bottom of the pond during flooding events and only those that are inadvertently swept out are able to leave. Every effort will be made to avoid pre-programmed releases including transfer to alternate facilities. Emergency releases, if necessary and authorized, would be managed by removal of outlet screens and pull sumps of the rearing units. If possible, staff would set up portable pumps to use river water to flush the fish.

10.11 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish resulting from fish releases.

- All program fish are mass marked for easy identification. Returning hatchery fish are under heavy selective harvest and are identified by an adipose fin-clip. Recycling downstream for sport harvest opportunity eliminates as many fish as possible removing potential spawners.
- The production and release of only smolts through fish culture and release practices fosters rapid seaward migration with minimal delay in the rivers, limiting interactions with naturally-produced steelhead juveniles.
- Release is from a location downstream of much of the habitat of listed Chinook and steelhead.
- Releases from this program have been shown not to stray outside the lower Columbia (Marston and Iverson 2012).
- WDFW fish health and operational concerns for Kalama Falls Hatchery programs are communicated to WDFW Region 5 staff for any risk management or needed treatment. See also HGMP section 9.7.
- WDFW proposes to continue monitoring, research and reporting of hatchery smolt migration performance behavior, and intra and interspecific interactions with wild fish to access, and adjust if necessary, hatchery production and release strategies to minimize effects on wild fish.

11 SECTION 11. MONITORING AND EVALUATION OF PERFORMANCE INDICATORS

11.1 Monitoring and evaluation of "Performance Indicators" presented in Section 1.10.

11.1.1 Describe plans and methods proposed to collect data necessary to respond to each "Performance Indicator" identified for the program.

Performance indicators for harvest will be accomplished by continuing mass marking (ad clip). CWT recoveries will help determine stray rate contributions on spawning grounds by watersheds close in proximity to this program's release vicinity. See HGMP section 1.10 Monitoring and Evaluation for additional plans and methods to collect data necessary.

Additional research, monitoring and evaluation in the Lower Columbia. WDFW is currently conducting the following Mitchell Act-funded research, monitoring and evaluation projects:

Table 11.1.1: Current WDFW Mitchell Act-funded research, monitoring and evaluation projects.

Project	Description
LCR Monitoring	WDFW has implemented an expanded monitoring program for Chinook, coho, chum and steelhead populations in the Lower Columbia River (LCR) region of Southwest Washington (WDFW's Region 5) and fishery monitoring in the lower mainstem of the Columbia River. The focus of this expanded monitoring is to 1) gather data on Viable Salmonid Population (VSP) parameters – spawner abundance, including proportion of hatchery-origin spawners (pHOS), spatial distribution, diversity, and productivity, 2) to increase the coded wire tag (CWT) recovery rate from spawning grounds to meet regional standards, and 3) to evaluate the use of PIT tags to develop harvest rates for salmon and steelhead populations. Additionally, key watersheds are monitored for juvenile

	salmonid out-migrant abundance. Coupled with adult abundance information, these data sets allow for evaluation of freshwater productivity and development of biological reference points, such as seeding capacity. Monitoring protocols and analysis methods utilized are intended to produce unbiased estimates with measurements of precision in an effort to meet NOAA monitoring guidelines (Crawford and Rumsey 2011).

11.1.2 Indicate whether funding, staffing, and other support logistics are available or committed to allow implementation of the monitoring and evaluation program.

Except for a risk involving genetic introgression, all other aspects of the M&E outlined in HGMP section 1.10 are currently funded (see also HGMP section 11.1.1).

11.2 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish resulting from monitoring and evaluation activities.

Monitoring, evaluation and research follow scientific protocols with adaptive management process if needed. WDFW will take risk aversion measures to eliminate or reduce ecological effects, injury, or mortality as a result of monitoring activities See HGMP section 1.10 Monitoring and Evaluation for additional plans and methods to collect data necessary, In addition, we will adaptively manage all aspects of the program to continue to minimize associated risks using the more recent available scientific research.

Trap box conditions. The highest potential for injury or mortality will likely be due to overloading of the trap box, which could be exacerbated by low flow/warm water conditions. To minimize this, the trap will be staffed nearly continuously while installed and the trap box will be checked multiple times/day, as necessary; temporary holding time will be <24 hours. In addition, water temperature will be monitored. If abundance of salmonids exceeds the ability of staff to efficiently work through fish, modifications to the sampling schedule and/or trapping protocols will be made to facilitate passage without handling. At all locations, this can be accomplished by opening the upstream gate on the trap box and allowing fish to pass through without handling, or by removing (or submerging) a panel section of the weir to allow fish passage around the trap box.

High flow events could prevent access to the trap box and limit WDFW staff's ability to handle fish, potentially trapping fish for the duration of the high flow event. WDFW project staff will monitor stream gauges to gather near real-time information on streamflows throughout the Lower Columbia. Gauges are operated by Washington Department of Ecology (DOE) on the Washougal River (station ID: 28B080). Utilizing streamflow, weather forecast information, and direct observation, WDFW personnel will determine when flows begin to limit the ability to access the trap box and sample fish. If these conditions are encountered the trap box will either be 1) opened on both the upstream and downstream end to allow direct passage through the trap, or 2) closed on both the upstream and downstream ends to prevent fish from becoming entrapped while personnel cannot access the trap. Which option is chosen will depend on the extent of the high flow event, expected duration, and the trap counts (i.e. relative abundance of fish that may be impeded) in the weekly trapping period prior to the event.

Fish handling and tagging. Handling and tagging of fish presents another potential injury/mortality risk. To minimize this, experienced, senior level staff will be overseeing handling and operations and ensuring field technicians are well trained in proper fish handling techniques. In addition, anesthetic may be used to calm fish during intrusive tagging procedures.

Impeding fish movement. Close attention will be paid to the recruitment of fish into the adult trap and the accumulation of fish below the trap. If fish are not adequately moving into the trap, modifications will first be made to adjust flow and try to increase trapping efficiency. If this does not encourage fish to move into the live box, a beach seine will be used to either capture fish or crowd them into the live box. The final option if fish are stacking up below the weir and cannot be captured through trapping or seining will be to remove or submerge weir panels to allow fish passage upstream of the weir for short intervals.

Spawning ground surveys and biological sampling occurring during the recovery will employ measures to ensure that effects on the survival of the listed Chinook salmon population are insignificant. Salmon redds and live spawning fish will not be disturbed during surveys and sampling.

12 SECTION 12. RESEARCH

12.1 Objective or purpose.

No research is directly associated with the program.

12.2 Cooperating and funding agencies.

Any research is conducted by WDFW and funded through Tacoma Power.

12.3 Principle investigator or project supervisor and staff.

Cowlitz Hatchery Biologist

12.4 Status of stock, particularly the group affected by project, if different than the stock(s) described in Section 2.

Not applicable.

12.5 Techniques: include capture methods, drugs, samples collected, tags applied.

Not applicable.

12.6 Dates or time period in which research activity occurs.

Not applicable.

12.7 Care and maintenance of live fish or eggs, holding duration, transport methods.

Not applicable.

12.8 Expected type and effects of take and potential for injury or mortality.

Not applicable.

12.9 Level of take of listed fish: number or range of fish handled, injured, or killed by sex, age, or size, if not already indicated in Section 2 and the attached "take table" (Table 1).

Not applicable.

12.10 Alternative methods to achieve project objectives.

Not applicable.

12.11 List species similar or related to the threatened species; provide number and causes of mortality related to this research project.

Not applicable.

12.12 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse ecological effects, injury, or mortality to listed fish as a result of the proposed research activities.

Not applicable.

13 SECTION 13. ATTACHMENTS AND CITATIONS

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Attachment 1: Fish Health Summaries: Kalama Hatchery Complex, April 1, 2007 through September 30, 2007 to October 1, 2011 through March 31, 2012.

Fallert Creek Hatchery Chinook

Juveniles: fall Chinook

2006 brood fall Chinook

Bacterial Gill Disease (BGD) in two raceways was detected in May 2007; fish were treated with hydrogen peroxide. It was observed that fish started on the EWOS starter feed had nutritional BGD and dropout syndrome; it was concluded that EWOS was not digested well in cold and fluctuating temperatures, and there was too great a size difference between #0 and #1, causing a poor transition between feeds. The fish grew much better on BioVita.

Water temperatures and disease continued to hamper growth; the fish were continuously sick with *Furunculosis* or *Icthyobodiasis*, and were treated with Paracide-S, oxytetracycline, and Romet. Fish were released in late-June/early-July 2007.

2007 brood fall Chinook

Fry were ponded and no significant loss was experienced. They grew very well on BioVita. Fish remained healthy, with excellent fat storage and condition factors.

2008 brood year fall Chinook

Fry were ponded with no significant loss. BGD was diagnosed in April 2009; fish were treated with hydrogen peroxide for three days, with minimal loss. Fish were released throughout June 2009, with a final release in July 2009.

2009 brood year fall Chinook (mixed)

Eyed-eggs (2.8-million) were received from Kalama Falls Hatchery in November and December 2009. These fish did doing well in their initial rearing period, and remained healthy through their release in June 2010.

2010 brood year fall Chinook (mixed)

Green eggs were taken at Fallert Creek Hatchery and Kalama Falls Hatchery. Eyed-eggs were also transferred from KFM; unfed fry were also received from KFM in March. Fry were ponded and no significant loss was experienced. Fish were growing well until April, when high turbidity of the water supply caused feeding problems, increasing the amount of drop out. In May 2011, the fish were diagnosed with *Ichthyobodo* and bacterial gill disease (BGD). The fish were initially treated hydrogen peroxide for the BGD and then formalin to treat the *Ichthyobodo*. These treatments had to be repeated in early-June on one raceway. Some level of dropout continued until fish were released in June and July.

2011 brood year fall Chinook – Kalama stock

Unfed fry were transferred from Kalama Falls Hatchery in late-March 2012. This stock remained healthy until May, then two of the five raceways began having problems with *Ichthyobodo,* requiring three separate formalin treatments. In late-May the fish in one of these raceways began dying from *Furunculosis* , and again in four of the five raceways at the end of June; the fish were treated with Romet® both times. The larger fish that had been moved into the rearing ponds suffered from lower levels of loss and were not treated. Fish were released early-June through early-July.

2012 brood year fall Chinook – Kalama stock

This stock remained healthy since ponding, except for a low level loss due to gut fungus (caused by *Phoma*), until the end of April 2013, when they went through a period of dropout. In mid-May 2013, they

suffered from an *Ichthyobodia* infestation which was treated with a 1:6,000 formalin drip treatment. The group of fish that was transferred from KFH had problems with BGD prior to transfer, and were debilitated and initially suffered from a high level of dropout. All fish were healthy at release in early-June 2013.

Kalama Falls Hatchery Chinook

Juveniles: fall Chinook

2006 brood fall Chinook

Difficulties with the EWOS starter feed and delay in getting the fish all on BioVita resulted in lingering dropout syndrome. BCD was treated with hydrogen peroxide in late-April 2007. This subpopulation remained problematic throughout their rearing period. BCWD began in October 2007, and was treated with Aquaflor. *Ichthyophthirius* was found in November, and fish were very out of size. Fish were transferred to Gobar Pond in December 2007. Predation was higher than normal due to a family of otters that moved in over the winter. Releases took place March 2008.

2006 brood fall Chinook

Feed-related problems with the EWOS starter feed that resulted in dropout syndrome. There were also significant problems with BGD, which was treated with hydrogen peroxide.

2007 brood fall Chinook

Egg-take goal of 5.5-million was not met in fall 2007. Due to colder water temperatures, these fish were ponded a few weeks later than normal and early-growth rates were slow. Fish were given BioVita feed, and combined lower densities and high quality food resulted in low mortalities, good growth conversion and excellent CVs. There were no fish health issues and they were released late-June through early-July 2008.

2008 brood year fall Chinook

Egg-takes began late-September 2008. The fall Chinook program releases was increased from 5-million to 7-million; egg-take in 2008 was only 6.2-million. In April 2009, low levels of *Ichthyobodo* were detected on the gills and the fry were also reportedly off their feed. This was remedied with a formalin treatment. Rearing mortalities were low, until early-May, with higher rearing densities. In June, dissolved oxygen levels (D.O. at 6.5 ppm or less) continued to be an issue with the increase in densities; North Toutle Hatchery was able to loan KFM seven aerators. The fish were healthy at release mid-June through early-July 2009.

2009 brood fall Chinook – Kalama River stock

The first egg-take occurred mid-September 2009. Eyed-eggs (2.8-million) were transferred to Fallert Creek Hatchery. Fish were ponded during January, February and March.

Systemic BCWD has been found in these fish; they received Aquaflor medicated feed. Mass-marking was delayed for one week to allow for the treatment. The loss level does not appear to exceed 0.1%, but turbid water and high flows prevented a complete estimate of loss. There was a light infestation of *Ichthyobodo* on the gills in July 2010; there was no need for treatment.

Rearing densities and dissolved oxygen levels were a challenge. Use of aerators and shipping a portion to Fallert Creek Hatchery for release helped. They were released healthy in mid-June 2010.

2010 brood year fall Chinook (mixed)

Egg-take began in mid-September 2010. Unfed fry were shipped to Fallert Creek Hatchery in February 2011. Fish remaining at Kalama Falls Hatchery were ponded in January and February, and grew well (on Biovita) until March. In March 2011, there was an increase in the number of pinheads, fish were not feeding well and were flashing in the pond. At this time they were diagnosed with a *Costia* infestation and were treated with Paracide-S (formalin).

Fish went through a period of moderate loss due to dropout in May, but the general population remained healthy. At the end of June, they were diagnosed with low levels of *Ichthyobodo* again and were treated with formalin. The fish were healthy at release in late-June and early-July 2011.

2011 brood year fall Chinook

Egg-take began in late-September 2011. A total of 106,600 fry were killed during incubation due to water supply issue. Fish were ponded in February and March 2012, and were growing and doing well. Fish were healthy until the second week in April, when they suffered a low level loss due to *Ichthyobodo* which was treated with formalin. Clipping was started on these fish the fourth week in April. Loss was not high in most raceways but *Ichthyobodo* was present. Bacterial Gill Disease was present one of the raceways; these fish in raceway were treated with three hydrogen peroxide drip treatments on alternate days. The remaining raceways were treated with one formalin drip treatment to control the *Ichthyobodo*. Ichthyobodo was again present in early June and another formalin drip treatment was given to these fish. The fish were healthy until their release the end of June through mid-July 2012.

2012 brood year fall Chinook – Kalama stock

These fish remained healthy until March 2013, when they began suffering from Bacterial Gill Disease. The fish were treated with hydrogen peroxide, but they continued to suffer from dropout loss throughout April requiring a number of hydrogen peroxide treatments. The fish were being marked in April, and raceways had reached maximum loadings; worked with crew to reevaluate feeding practices such as reducing feeding levels and getting the feed into the fish in less time rather than hourly feedings throughout the day. In May 2013, losses continued due to dropout, which was higher than normal due to the earlier BGD problems. At the end of May the fish had light *Ichthyobodo* present but it did not warrant treatment. The fish remained healthy until their release in June 2013.

Attachment 2: WDFW Virology Sampling 2006-2007 through 2012-13: Kalama Falls and Fallert Creek Hatcheries Chinook.

Hatchery/ Collection Site	Stock	Species	Date Sampled	Results	Comments	Life Stage	Sample number	Number of fish sampled						ID	Cell Line	Frozen	Inoc Date
								OF	pools	K/S	pools	fry/visc	pools				
KALAMA FLS	KALAMA R	FCHIN	10/03/06	NEV		AD	1003-11/12	60	12			60	12				
KALAMA FLS	KALAMA R	SPCHIN	09/12/06	NEV		AD	0913-5/6	24	5			24	5				
KALAMA FLS	KALAMA R	SPCHIN	09/19/06	NEV		AD	0920-12/13	36	8			36	8				
KALAMA FLS	KALAMA R	SPCHIN	09/05/07	NEV		AD	0907-1/2	60	12			60	12				
KALAMA FLS	KALAMA R	FCHIN	10/08/07	NEV		AD	1009-9/10	55	11			60	12				
KALAMA FLS	KALAMA R	FCHIN	10/17/07	NEV		AD	1018-5	5	1								
KALAMA FLS	KALAMA R	SPCHIN	09/09/08	NEV		AD	0910-5/6	60	12			60	12				
KALAMA FLS	KALAMA R	FCHIN	09/30/08	NEV		AD	1001-7/8	60	12			60	12				
KALAMA FLS	KALAMA R	SPCHIN	09/01/09	NEV		AD	0902-7/8	15	3			15	3				
KALAMA FLS	KALAMA R	SPCHIN	09/01/09	NEV		AD	0902-7/8	15	3			15	3				
KALAMA FLS	KALAMA R	SPCHIN	09/09/09	NEV		AD	0910-8/9	20	5			21	5				
KALAMA FLS	KALAMA R	SPCHIN	09/09/09	NEV		AD	0910-8/9	20	5			21	5				
KALAMA FLS	KALAMA R	SPCHIN	09/15/09	NEV		AD	0916-1/2	5	1			10	2				
KALAMA FLS	KALAMA R	SPCHIN	09/15/09	NEV		AD	0916-1/2	5	1			10	2				
KALAMA FLS	KALAMA R	SPCHIN	09/21/09	NEV		AD	0922-8/9	1	1			1	1				
KALAMA FLS	KALAMA R	SPCHIN	09/21/09	NEV		AD	0922-8/9	1	1			1	1				
KALAMA FLS	KALAMA R	FCHIN	09/22/09	NEV		AD	0922-20					10	10				
KALAMA FLS	KALAMA R	FCHIN	09/22/09	NEV		AD	0922-20					10	10				
KALAMA FLS	KALAMA R	FCHIN	09/29/09	**IHNV**	1+/12p K/S	AD	0929-11/12	60	12			60	12		E/C	10/28/09	
KALAMA FLS	KALAMA R	FCHIN	09/29/09	**IHNV**	1+/12p K/S	AD	0929-11/12	60	12			60	12		E/C	10/28/09	
KALAMA FLS	KALAMA R	FCHIN	10/06/09	NEV	pond 13-14	AD	1006-11	12	6								
KALAMA FLS	KALAMA R	FCHIN	10/06/09	NEV	pond 13-14	AD	1006-11	12	6								
FALLERT CR	KALAMA R	FCHIN	10/07/09	NEV		AD	1007-3/4	60	12			60	12				
FALLERT CR	KALAMA R	FCHIN	10/07/09	NEV		AD	1007-3/4	60	12			60	12				
KALAMA FLS	KALAMA R	SPCHIN	09/14/10	NEV		AD	0914-7/8	53	11			60	12				
KALAMA FLS	KALAMA R	SPCHIN	09/20/10	NEV		AD	0921-16	10	2								
KALAMA FLS	KALAMA R	FCHIN	10/06/10	NEV		AD	1006-13/14	60	12			60	12				
KALAMA FLS	KALAMA R	SPCHIN	09/13/11	**IHNV**	OF: 1+/12P	AD	0914-1/2	60	12			60	12	S/N		11/22/11	
FALLERT	KALAMA R	FCHIN	10/11/11	NEV		AD	1012-8/9	40	8			50	10				
FALLERT	KALAMA R	FCHIN	10/19/11	NEV		AD	1020-3/4	20	4			10	2				
KALAMA FLS	KALAMA R	FCHIN	10/20/11	**IHNV**	OF and K/S: 1+/12P	AD	1021-1/2	60	12			60	12	DB		11/10/11	
KALAMA FLS	KALAMA R	SPCHIN	08/29/12	NEV		AD	0830-5/6	7	2	12	3						12/27/12
KALAMA FLS	KALAMA R	SPCHIN	09/05/12	NEV		AD	0906-2/3	24	5	24	5						11/7/12 & 11/8/12
KALAMA FLS	KALAMA R	SPCHIN	09/11/12	NEV		AD	0912-1/2	30	6	25	5						2/21/13
FALLERT CR	KALAMA R	SPCHIN	09/26/12	NEV	DIAG EPC 10E0-10E-3	JUV/11	0927-1			3	1						1/3/13
FALLERT CR	KALAMA R	FCHIN	10/09/12	NEV		AD	1010-1/2	60	12	60	12						9/11/12
KALAMA FLS	KALAMA R	FCHIN	10/10/12	NEV		AD	1011-2/3	60	12	60	12						11/27/12
KALAMA FLS	COWLITZ R	SPCHIN	03/04/13	NEV	Raceway 11 EPC 10-0 to 10-3 All alive until processed	JUV 12	0305-1					6	2				9/26/12

Source: WDFW Fish Health Lab data 2014 (John Kerwin).

Note: For Kalama system steelhead data, see Kalama summer and winter steelhead HGMPs.

14 SECTION 14. CERTIFICATION LANGUAGE AND SIGNATURE OF RESPONSIBLE PARTY

"I hereby certify that the information provided is complete, true and correct to the best of my knowledge and belief. I understand that the information provided in this HGMP is submitted for the purpose of receiving limits from take prohibitions specified under the Endangered Species Act of 1973 (16 U.S.C.1531-1543) and regulations promulgated thereafter for the proposed hatchery program, and that any false statement may subject me to the criminal penalties of 18 U.S.C. 1001, or penalties provided under the Endangered Species Act of 1973."

Name, Title, and Signature of Applicant:

Certified by_____ Date:_____

15 ADDENDUM A. PROGRAM EFFECTS ON OTHER (AQUATIC OR TERRESTRIAL) ESA-LISTED POPULATIONS. (Anadromous salmonid effects are addressed in Section 2).

15.1 List all ESA permits or authorizations for USFWS ESA-listed, proposed, and candidate salmonid and non-salmonid species associated with the hatchery program.

The WDFW and the USFWS have a Cooperative Agreement pursuant to section 6(c) of the Endangered Species Act that covers the majority of the WDFW actions, including hatchery operations.

> *"The department is authorized by the USFWS for certain activities that may result in the take of bull trout, including salmon/steelhead hatchery broodstocking, hatchery monitoring and evaluation activities and conservation activities such as adult traps, juvenile monitoring, spawning ground surveys..."*

15.2 Describe USFWS ESA-listed, proposed, and candidate salmonid and non-salmonid species and habitat that may be affected by hatchery program.

Several USFWS listed and candidate species are found in Cowlitz County, however the hatchery operations and facilities for this program do not fall within the critical habitat for any of these species. As such there are no effects anticipated for these species.

Listed or candidate species:
"No effect" for the following species:

Bull trout (*Salvelinus confluentus*) – Threatened (Critical Habitat Designated)
Nelson's checker-mallow (*Sidalcea nelsoniana*) –Threatened
Marbled murrelet (*Brachyramphus marmoratus*) –Threatened (Critical Habitat Designated)
Columbian White-Tailed deer (*Odocoileus virginianus leucurus*) – Endangered
Gray Wolf (*Canis lupus*) –Threatened
Northern Spotted owl (*Strix occidentalis caurina*) –Threatened (Critical Habitat Designated)

Candidate Species
North American wolverine *(Gulo gulo luteus)* – contiguous U.S. DPS

15.3 Analyze effects.

Not applicable.

15.4 Actions taken to minimize potential effects.

Program coho are released fully smolted to foster rapid outmigration from the basin and to minimize predation and residualism risks.

15.5 References

Not applicable.

16 "Take" Tables

Table 1. Estimated listed salmonid take levels of by hatchery activity.

Listed species affected: Spring Chinook (*Oncorhynchus tshawytscha*)	**ESU/Population:** Lower Columbia River Spring Chinook	**Activity:** Kalama Fall Chinook Program
Location of hatchery activity: Kalama Falls Hatchery, Kalama River (WRIA 27.0002)at RKm 16.1. Fallert Creek Hatchery, Fallert Creek (WRIA 27.0017) at RKm 8.2; tributary to the Kalama River at RKm. 4.9. Modrow Trap, Kalama River at RKm 4.8.	**Dates of activity:** August-October	**Hatchery program operator:** WDFW

Type of Take	**Annual Take of Listed Fish By Life Stage *(Number of Fish)***			
	Egg/Fry	**Juvenile/Smolt**	**Adult**	**Carcass**
Observe or harass [a]				
Collect for transport [b]				
Capture, handle, and release [c]			TBD	
Capture, handle, tag/mark/tissue sample, and released [d]				
Removal (e.g. broodstock) [e]				
Intentional lethal take [f]				
Unintentional lethal take [g]		TBD	TBD	
Other Take (specify) [h]				

Take Tables to be submitted to NOAA-NMFS, in progress.

a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
e. Listed fish removed from the wild and collected for use as broodstock.
f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
h. Other takes not identified above as a category.

Instructions:

1. *An entry for a fish to be taken should be in the take category that describes the greatest impact.*
2. *Each take to be entered in the table should be in one take category only (there should not be more than one entry for the same sampling event).*
3. *If an individual fish is to be taken more than once on separate occasions, each take must be entered in the take table.*

Table 2. Estimated listed salmonid take levels of by hatchery activity.

Listed species affected: Fall Chinook (*Oncorhynchus tshawytscha*)	**ESU/Population:** Lower Columbia River Fall Chinook	**Activity:** Kalama Fall Chinook Program
Location of hatchery activity: Kalama Falls Hatchery, Kalama River (WRIA 27.0002)at RKm 16.1. Fallert Creek Hatchery, Fallert Creek (WRIA 27.0017) at RKm 8.2; tributary to the Kalama River at RKm. 4.9. Modrow Trap, Kalama River at RKm 4.8.	**Dates of activity:** August-October	**Hatchery program operator:** WDFW

Type of Take	**Annual Take of Listed Fish By Life Stage *(Number of Fish)***			
	Egg/Fry	**Juvenile/Smolt**	**Adult**	**Carcass**
Observe or harass [a]				
Collect for transport [b]				
Capture, handle, and release [c]				
Capture, handle, tag/mark/tissue sample, and released [d]		TBD		
Removal (e.g. broodstock) [e]			TBD	
Intentional lethal take [f]		TBD		
Unintentional lethal take [g]		TBD		
Other Take (specify) [h]				

a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
e. Listed fish removed from the wild and collected for use as broodstock.
f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
h. Other takes not identified above as a category.

Table 3. Estimated listed salmonid take levels of by hatchery activity.

Listed species affected: Steelhead *(Oncorhynchus mykiss*)	**ESU/Population:** Lower Columbia River Steelhead	**Activity:** Kalama Fall Chinook Program
Location of hatchery activity: Kalama Falls Hatchery, Kalama River (WRIA 27.0002)at RKm 16.1 Fallert Creek Hatchery, Fallert Creek (WRIA 27.0017) at RKm 8.2; tributary to the Kalama River at RKm. 4.9. Modrow Trap, Kalama River at RKm 4.8.	**Dates of activity:** September-January	**Hatchery program operator:** WDFW

Type of Take	**Annual Take of Listed Fish By Life Stage *(Number of Fish)***			
	Egg/Fry	**Juvenile/Smolt**	**Adult**	**Carcass**
Observe or harass [a]				
Collect for transport [b]				
Capture, handle, and release [c]			TBD	
Capture, handle, tag/mark/tissue sample, and released [d]				
Removal (e.g. broodstock) [e]				
Intentional lethal take [f]				
Unintentional lethal take [g]		TBD	TBD	
Other Take (specify) [h]				

a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
e. Listed fish removed from the wild and collected for use as broodstock.
f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
h. Other takes not identified above as a category.

Table 4. Estimated listed salmonid take levels of by hatchery activity.

Listed species affected: Coho (*Oncorhynchus kisutch*)	**ESU/Population:** Lower Columbia River Coho	**Activity:** Kalama Fall Chinook Program
Location of hatchery activity: Kalama Falls Hatchery, Kalama River (WRIA 27.0002)at RKm 16.1 Fallert Creek Hatchery, Fallert Creek (WRIA 27.0017) at RKm 8.2; tributary to the Kalama River at RKm. 4.9. Modrow Trap, Kalama River at RKm 4.8.	**Dates of activity:** August-October	**Hatchery program operator:** WDFW

Type of Take	**Annual Take of Listed Fish By Life Stage *(Number of Fish)***			
	Egg/Fry	**Juvenile/Smolt**	**Adult**	**Carcass**
Observe or harass [a]				
Collect for transport [b]				
Capture, handle, and release [c]			TBD	
Capture, handle, tag/mark/tissue sample, and released [d]				
Removal (e.g. broodstock) [e]				
Intentional lethal take [f]				
Unintentional lethal take [g]		TBD	TBD	
Other Take (specify) [h]				

a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
e. Listed fish removed from the wild and collected for use as broodstock.
f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
h. Other takes not identified above as a category.